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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 15/57, 9/64, 9/48, 1/19, 5/10, 1/21, A61K 38/48, C07K 16/40	A3	(11) International Publication Number: WO 99/36550 (43) International Publication Date: 22 July 1999 (22.07.99)
(21) International Application Number: PCT/US99/00655 (22) International Filing Date: 12 January 1999 (12.01.99) (30) Priority Data: 09/008,271 16 January 1998 (16.01.98) US (63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application US 09/008,271 (CIP) Filed on 16 January 1998 (16.01.98) (71) Applicant (for all designated States except US): INCYTE PHARMACEUTICALS, INC. [US/US]; 3174 Porter Drive, Palo Alto, CA 94304 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): BANDMAN, Olga [US/US]; 366 Anna Avenue, Mountain View, CA 94043 (US). HILLMAN, Jennifer, L. [US/US]; 230 Monroe Drive, #12, Mountain View, CA 94040 (US). YUE, Henry [US/US]; 826 Lois Avenue, Sunnyvale, CA 94087 (US). GUEGLER, Karl, J. [CH/US]; 1048 Oakland Avenue, Menlo Park, Ca 94025 (US). CORLEY, Neil, C. [US/US]; 1240 Dale Avenue #30, Mountain View, CA 94040 (US).	TANG, Y., Tom [CN/US]; 110 East Remington Drive #14, Sunnyvale, CA 94087 (US). SHAH, Purvi [IN/US]; 1608 Queen Charlotte Drive #5, Sunnyvale, CA 94087 (US). (74) Agents: BILLINGS, Lucy, J. et al.; Incyte Pharmaceuticals, Inc., 3174 Porter Drive, Palo Alto, CA 94304 (US). (81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i> (88) Date of publication of the international search report: 18 November 1999 (18.11.99)	
(54) Title: HUMAN PROTEASE MOLECULES		
(57) Abstract The invention provides human protease molecules (HUPM) and polynucleotides which identify and encode HUPM. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for treating or preventing disorders associated with expression of HUPM.		

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INTERNATIONAL SEARCH REPORT

International Application No

PC1/US 99/00655

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/57 C12N9/64 C12N9/48 C12N1/19 C12N5/10
C12N1/21 A61K38/48 C07K16/40

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EMBL/GENBANK DATABASES Accession no T29740, Sequence reference HS74011 9 January 1995 ADAMS M ET AL: "Initial assessment of human gene diversity and expression patterns based upon 52 million basepairs of cDNA sequence" XP002103800 the whole document	1-13,15, 22,23
A	TAMURA T ET AL: "Molecular cloning and sequence analysis of cDNAs for five major subunits of human proteasomes (multi-catalytic proteinase complexes)" BIOCHIMICA ET BIOPHYSICA ACTA, vol. 1089, 1991, pages 95-102, XP002103799 figures 1D,2	

☒ Further documents are listed in the continuation of box C.☐ Patent family members are listed in annex.

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O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

Z document member of the same patent family

Date of the actual completion of the international search

26 May 1999

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>EMBL/GENBANK DATABASES Accession no P56202, Sequence reference LYMP_HUMAN 1 November 1997 BROWN J ET AL: "Lymphopain precursor" XP002103801 the whole document -----</p>	

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 99/ 00655

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claims 18-21
are directed to a method of treatment of the human/animal
body, the search has been carried out and based on the alleged
effects of the compound/composition.
2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such
an extent that no meaningful International Search can be carried out, specifically:
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all
searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment
of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report
covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is
restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-23 partially

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

1. Claims: Invention 1: claims 1-23 partially

Human protease molecule with amino acid sequence SEQ ID NO 1, nucleotides encoding the protease, antibodies against the enzyme, antagonists or agonists of the enzyme and their use

2. Claims: Inventions 2-12, claims 1-23 partially

Invention 2 being a human protease molecule with amino acid sequence SEQ ID NO 2, nucleotides encoding the protease, antibodies against the enzyme, antagonists or agonists of the enzyme and their use; Invention 3 being a human protease molecule with amino acid sequence SEQ ID NO 3, nucleotides encoding the protease, antibodies against the enzyme, antagonists or agonists of the enzyme and their use etc...
Invention 12 being a human protease molecule with amino acid sequence SEQ ID NO 12, nucleotides encoding the protease, antibodies against the enzyme, antagonists or agonists of the enzyme and their use etc...

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HUMAN PROTEASE MOLECULES

TECHNICAL FIELD

5 This invention relates to nucleic acid and amino acid sequences of human protease molecules and to the use of these sequences in the diagnosis, treatment, and prevention of cell proliferative and immune disorders.

BACKGROUND OF THE INVENTION

10 Proteolytic processing is an essential component of normal cell growth, differentiation, remodeling, and homeostasis. The cleavage of peptide bonds within cells is necessary for the maturation of precursor proteins to their active form, the removal of signal sequences from targeted proteins, the degradation of incorrectly folded proteins, and the controlled turnover of peptides within the cell. Proteases participate in apoptosis,
15 inflammation, and in tissue remodeling during embryonic development, wound healing, and normal growth. They are necessary components of bacterial, parasitic, and viral invasion and replication within a host. Four principal categories of mammalian proteases have been identified based on active site structure, mechanism of action, and overall three-dimensional structure. (Beynon, R.J. and J.S. Bond (1994) Proteolytic Enzymes: A
20 Practical Approach, Oxford University Press, New York, NY, pp. 1-5.)

 The serine proteases (SPs) are a large family of proteolytic enzymes that include the digestive enzymes, trypsin and chymotrypsin; components of the complement cascade and of the blood-clotting cascade; and enzymes that control the degradation and turnover of macromolecules of the extracellular matrix. SPs are so named because of the presence
25 of a serine residue found in the active catalytic site for protein cleavage and usually within the sequence GDSGGP. The active site of all SP is composed of a triad of residues including the aforementioned serine, an aspartate, and a histidine residue. SPs have a wide range of substrate specificities and can be subdivided into subfamilies on the basis of these specificities. The main sub-families are trypases which cleave after arginine or
30 lysine; aspases which cleave after aspartate; chymases which cleave after phenylalanine or leucine; metases which cleavage after methionine; and serases which cleave after serine.

The SPs are secretory proteins containing N-terminal signal peptides which export the immature protein across the endoplasmic reticulum prior to cleavage. (von Heijne, G. (1986) *Nuc. Acid. Res.* 14:5683-5690). Differences in these signal sequences provide one means of distinguishing individual SPs. Some SPs, particularly the digestive enzymes, exist as inactive precursors or preproenzymes and contain a leader or activation peptide on the C-terminal side of the signal peptide. This activation peptide may be 2-12 amino acids in length, and extend from the cleavage site of the signal peptide to the N-terminus of the active, mature protein. Cleavage of this sequence activates the enzyme. This sequence varies in different SPs according to the biochemical pathway and/or its substrate. (Zunino, S.J. et al. (1990) *J. Immunol.* 144:2001-2009; and Sayers, T.J. et al. (1994) *J. Immunol.* 152:2289-2297.)

Cysteine proteases are involved in diverse cellular processes ranging from the processing of precursor proteins to intracellular degradation. Mammalian cysteine proteases include lysosomal cathepsins and cytosolic calcium activated proteases, calpains. Cysteine proteases are produced by monocytes, macrophages and other cells of the immune system which migrate to sites of inflammation and in their protective role secrete various molecules to repair damaged tissue. These cells may overproduce the same molecules and cause tissue destruction in certain disorders. In autoimmune diseases such as rheumatoid arthritis, the secretion of the cysteine protease, cathepsin C, degrades collagen, laminin, elastin and other structural proteins found in the extracellular matrix of bones. The cathepsin family of lysosomal proteases includes the cysteine proteases; cathepsins B, H, K, L, O2, and S; and the aspartyl proteases; cathepsins D and G. Various members of this endosomal protease family are differentially expressed. Some, such as cathepsin D, have a ubiquitous tissue distribution while others, such as cathepsin L, are found only in monocytes, macrophages, and other cells of the immune system.

Abnormal regulation and expression of cathepsins is evident in various inflammatory disease states. In cells isolated from inflamed synovia, the mRNA for stromelysin, cytokines, TIMP-1, cathepsin, gelatinase, and other molecules is preferentially expressed. Expression of cathepsins L and D is elevated in synovial tissues from patients with rheumatoid arthritis and osteoarthritis. Cathepsin L expression may also contribute to the influx of mononuclear cells which exacerbates the destruction of the rheumatoid synovium. (Keyszer, G.M. (1995) *Arthritis Rheum.* 38:976-984.) The

increased expression and differential regulation of the cathepsins is linked to the metastatic potential of a variety of cancers and as such is of therapeutic and prognostic interest.

(Chambers, A.F. et al. (1993) Crit. Rev. Oncog. 4:95-114.)

Cysteine proteases are characterized by a catalytic domain containing a triad of
5 amino acid residues similar to that found in serine proteases. A cysteine replaces the active serine residue. Catalysis proceeds via a thiol ester intermediate and is facilitated by the side chains of the adjacent histidine and aspartate residues.

Aspartic proteases include bacterial penicillopepsin, mammalian pepsin, renin, chymosin, and certain fungal proteases. The characteristic active site residues of aspartic
10 proteases are a pair of aspartic acid residues, e.g., asp33 and asp213 in penicillopepsin. Aspartic proteases are also called acid proteases because the optimum pH for activity is between 2 and 3. In this pH range, one of the aspartate residues is ionized, the other un-ionized. A potent inhibitor of aspartic proteases is the hexapeptide, pepstatin, which in the transition state resembles normal substrates.

15 Carboxypeptidases A and B are the principal mammalian representatives of the metallo-protease family. Both are exopeptidases of similar structure and active sites. Carboxypeptidase A, like chymotrypsin, prefers C-terminal aromatic and aliphatic side chains of hydrophobic nature, whereas carboxypeptidase B is directed toward basic arginine and lysine residues. Active site components include zinc, with its three ligands of
20 two glutamic acid and one histidine residues.

Many other proteolytic enzymes do not fit any of the major categories discussed above because their mechanisms of action and/or active sites have not been elucidated. These include the aminopeptidases and signal peptidases.

Aminopeptidases catalyze the hydrolysis of amino acid residues from the amino
25 terminus of peptide substrates. Bovine leucine aminopeptidase is a zinc metallo-enzyme that utilizes the sulfhydryl groups from at least three reactive cysteine residues at its active site in the binding of metal ions. (Cuypers, H.T. et al. (1982) J. Biol. Chem. 257:7086-7091.)

Signal peptidases are a specialized class of proteases found in all prokaryotic and
30 eukaryotic cell types that serve in the processing of signal peptides from certain proteins. Signal peptides are amino-terminal sequences on a protein which directs the protein from its ribosomal assembly site to a particular cellular or extracellular location. Once the

protein has been exported, removal of the signal sequence by a signal peptidase and posttranslational processing, e.g., glycosylation or phosphorylation, activate the protein. Signal peptidases exist as multi-subunit complexes in both yeast and mammals. The canine signal peptidase complex is composed of five subunits; all associate with the
5 mitochondrial membrane, and containing hydrophobic regions that span the membrane one or more times. (Shelness, G.S. and Blobel, G. (1990) J. Biol. Chem. 265:9512-9519.) Some of these subunits serve to fix the complex in its proper position on the membrane while others contain the actual catalytic activity. The catalytic activity appears to involve a serine residue in its active site.

10 Proteasome is an intracellular protease complex which is found in some bacteria and in all eukaryotic cells and plays an important role in cellular physiology. Proteasomes are responsible for the timely degradation of cellular proteins of all types and control proteins that function to activate or repress cellular processes such as transcription and cell cycle progression. (Ciechanover, A. (1994) Cell 79:13-21.) Proteasomes act on proteins
15 which have been targeted for hydrolysis by the covalent attachment of a small protein called ubiquitin to lysine side chains of the protein. Ubiquitin-proteasome systems are implicated in the degradation of mitotic cyclic kinases, oncoproteins, tumor suppressor genes (p53), cell surface receptors associated with signal transduction, transcriptional regulators, and mutated or damaged proteins. (Ciechanover, *supra*.) Proteasomes are large
20 (~ 2000 kDa), multisubunit complexes composed of a central catalytic core containing a variety of proteases and terminal subunits that serve in substrate recognition and regulation of proteasome activity.

Protease inhibitors play a major role in the regulation of the activity and effect of proteases. They have been shown to control pathogenesis in animal models of proteolytic
25 disorders. (Murphy, G. (1991) Agents Actions Suppl 35:69-76.) In particular, low levels of the cystatins, low molecular weight inhibitors of the cysteine proteases, seem to be correlated with malignant progression of tumors. (Calkins, C. et al (1995) Biol Biochem Hoppe Seyler 376:71-80.) The balance between levels of cysteine proteases and their inhibitors is also significant in the development of disorders. Specifically, increases in
30 cysteine protease levels, when accompanied by reductions in inhibitor activity, are correlated with increased malignant properties of tumor cells and the pathology of arthritis and immunological diseases in humans.

The serpins are high molecular weight, e.g., 370-420 amino acid residues, inhibitors of mammalian plasma serine proteases. Many of these inhibitors serve to regulate the blood clotting cascade and/or the complement cascade in mammals. Prominent among these inhibitors are α -1 protease inhibitor, α -1-antichymotrypsin, antithrombin III, and the "universal protease inhibitor" α -2 macroglobulin. α -1 protease inhibitor is primarily effective against the neutrophil elastase but combines with other serine proteases as well. α -1 protease inhibitor, α -1-antichymotrypsin, and antithrombin III all show striking sequence homology, suggesting that specialization of these inhibitors has occurred in response to specialization of the corresponding proteases themselves.

10 The discovery of new human protease molecules and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, treatment, and prevention of cell proliferative and immune disorders.

SUMMARY OF THE INVENTION

15 The invention features substantially purified polypeptides, human protease molecules, referred to collectively as "HUPM" and individually as "HUPM-1", "HUPM-2", "HUPM-3", "HUPM-4", "HUPM-5", "HUPM-6", "HUPM-7", "HUPM-8", "HUPM-9", "HUPM-10", "HUPM-11", and "HUPM-12". In one aspect, the invention provides a substantially purified polypeptide, HUPM, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof.

The invention further provides a substantially purified variant of HUPM having at least 90% amino acid identity to the amino acid sequences of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, or to a fragment of any of these sequences. The invention also provides an isolated and purified polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof. The invention also includes an isolated and purified polynucleotide variant having at least 90%

polynucleotide identity to the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof.

Additionally, the invention provides a composition comprising a polynucleotide encoding the polypeptide comprising the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof. The invention further provides an isolated and purified polynucleotide sequence which hybridizes under stringent conditions to the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof, as well as an isolated and purified polynucleotide sequence which is complementary to the polynucleotide encoding the polypeptide comprising the amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof.

The invention also provides an isolated and purified polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24. The invention further provides an isolated and purified polynucleotide variant having at least 90% polynucleotide identity to the polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24, and fragments thereof, as well as an isolated and purified polynucleotide which is complementary to the polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ

ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24, and fragments thereof.

The invention further provides an expression vector containing at least a fragment of the polynucleotide encoding the polypeptide comprising an amino acid sequence
5 selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof. In another aspect, the expression vector is contained within a host cell.

The invention also provides a method for producing a polypeptide comprising the
10 amino acid sequence of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12, or fragments thereof, the method comprising the steps of: (a) culturing the host cell containing an expression vector containing at least a fragment of a polynucleotide sequence encoding HUPM under conditions suitable for the expression
15 of the polypeptide; and (b) recovering the polypeptide from the host cell culture.

The invention also provides a pharmaceutical composition comprising a substantially purified HUPM having the amino acid sequence of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12, or fragments
20 thereof, in conjunction with a suitable pharmaceutical carrier.

The invention further includes a purified antibody which binds to a polypeptide comprising the amino acid sequence of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12, or fragments thereof, as well as a
25 purified agonist and a purified antagonist to the polypeptide.

The invention also provides a method for treating or preventing a cell proliferative disorder associated with increased expression or activity of HUPM, the method comprising administering to a subject in need of such treatment an effective amount of an antagonist of HUPM.

30 The invention also provides a method for treating or preventing an immune disorder associated with increased expression or activity of HUPM, the method comprising administering to a subject in need of such treatment an effective amount of an

antagonist of HUPM.

The invention also provides a method for treating or preventing a cell proliferative disorder associated with decreased expression or activity of HUPM, the method comprising administering to a subject in need of such treatment an effective amount of a
5 pharmaceutical composition comprising HUPM in conjunction with a suitable pharmaceutical carrier.

The invention also provides a method for treating or preventing an immune disorder associated with decreased expression or activity of HUPM, the method comprising administering to a subject in need of such treatment an effective amount of a
10 pharmaceutical composition comprising HUPM in conjunction with a suitable pharmaceutical carrier.

The invention also provides a method for detecting a polynucleotide encoding HUPM in a biological sample containing nucleic acids, the method comprising the steps of: (a) hybridizing the complement of the polynucleotide sequence encoding the
15 polypeptide comprising SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, or SEQ ID NO:12, or fragments thereof to at least one of the nucleic acids of the biological sample, thereby forming a hybridization complex; and (b) detecting the hybridization complex, wherein the presence of the hybridization complex correlates with
20 the presence of a polynucleotide encoding HUPM in the biological sample. In one aspect, the nucleic acids of the biological sample are amplified by the polymerase chain reaction prior to the hybridizing step.

25

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular methodology, protocols, cell lines, vectors, and reagents described, as these may vary. It is also to be understood that
30 the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms “a,” “an,” and “the” include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to “a host cell” includes a plurality of such host cells, and a reference to “an antibody” is a reference to one or more antibodies and equivalents thereof
5 known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the
10 preferred methods, devices, and materials are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, vectors, and methodologies which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

15

DEFINITIONS

“HUPM,” as used herein, refers to the amino acid sequences of substantially purified HUPM obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and preferably the human species, from any
20 source, whether natural, synthetic, semi-synthetic, or recombinant.

The term “agonist,” as used herein, refers to a molecule which, when bound to HUPM, increases or prolongs the duration of the effect of HUPM. Agonists may include proteins, nucleic acids, carbohydrates, or any other molecules which bind to and modulate the effect of HUPM.

25 An “allele” or an “allelic sequence,” as these terms are used herein, is an alternative form of the gene encoding HUPM. Alleles may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. Any given natural or recombinant gene may have none, one, or many allelic forms. Common mutational changes which give
30 rise to alleles are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding HUPM, as described herein, include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polynucleotide the same HUPM or a polypeptide with at least one functional characteristic of HUPM. Included within this definition are polymorphisms which may or
5 may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding HUPM, and improper or unexpected hybridization to alleles, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding HUPM. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result
10 in a functionally equivalent HUPM. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of HUPM is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, positively charged amino acids may include lysine and arginine,
15 and amino acids with uncharged polar head groups having similar hydrophilicity values may include leucine, isoleucine, and valine; glycine and alanine; asparagine and glutamine; serine and threonine; and phenylalanine and tyrosine.

The terms "amino acid" or "amino acid sequence," as used herein, refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and
20 to naturally occurring or synthetic molecules. In this context, "fragments", "immunogenic fragments", or "antigenic fragments" refer to fragments of HUPM which are preferably about 5 to about 15 amino acids in length and which retain some biological activity or immunological activity of HUPM. Where "amino acid sequence" is recited herein to refer to an amino acid sequence of a naturally occurring protein molecule, "amino acid
25 sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification," as used herein, relates to the production of additional copies of a nucleic acid sequence. Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art. (See, e.g., Dieffenbach, C.W. and
30 G.S. Dveksler (1995) PCR Primer, a Laboratory Manual, Cold Spring Harbor Press, Plainview, NY, pp.1-5.)

The term "antagonist," as it is used herein, refers to a molecule which, when bound

to HUPM, decreases the amount or the duration of the effect of the biological or immunological activity of HUPM. Antagonists may include proteins, nucleic acids, carbohydrates, antibodies, or any other molecules which decrease the effect of HUPM.

As used herein, the term "antibody" refers to intact molecules as well as to
5 fragments thereof, such as Fa, F(ab')₂, and Fv fragments, which are capable of binding the epitopic determinant. Antibodies that bind HUPM polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized
10 chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant," as used herein, refers to that fragment of a
15 molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (given regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the
20 immune response) for binding to an antibody.

The term "antisense," as used herein, refers to any composition containing a nucleic acid sequence which is complementary to a specific nucleic acid sequence. The term "antisense strand" is used in reference to a nucleic acid strand that is complementary
to the "sense" strand. Antisense molecules may be produced by any method including
25 synthesis or transcription. Once introduced into a cell, the complementary nucleotides combine with natural sequences produced by the cell to form duplexes and to block either transcription or translation. The designation "negative" can refer to the antisense strand, and the designation "positive" can refer to the sense strand.

As used herein, the term "biologically active," refers to a protein having structural,
30 regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" refers to the capability of the natural, recombinant, or synthetic HUPM, or of any oligopeptide thereof, to induce a specific immune response in

appropriate animals or cells and to bind with specific antibodies.

The terms "complementary" or "complementarity," as used herein, refer to the natural binding of polynucleotides under permissive salt and temperature conditions by base pairing. For example, the sequence "A-G-T" binds to the complementary sequence
5 "T-C-A." Complementarity between two single-stranded molecules may be "partial," such that only some of the nucleic acids bind, or it may be "complete," such that total complementarity exists between the single stranded molecules. The degree of complementarity between nucleic acid strands has significant effects on the efficiency and strength of the hybridization between the nucleic acid strands. This is of particular
10 importance in amplification reactions, which depend upon binding between nucleic acids strands, and in the design and use of peptide nucleic acid (PNA) molecules.

A "composition comprising a given polynucleotide sequence" or a "composition comprising a given amino acid sequence," as these terms are used herein, refer broadly to any composition containing the given polynucleotide or amino acid sequence. The
15 composition may comprise a dry formulation, an aqueous solution, or a sterile composition. Compositions comprising polynucleotide sequences encoding HUPM or fragments of HUPM may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g.,
20 NaCl), detergents (e.g., SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

The phrase "consensus sequence," as used herein, refers to a nucleic acid sequence which has been resequenced to resolve uncalled bases, extended using XL-PCR™ (Perkin Elmer, Norwalk, CT) in the 5' and/or the 3' direction, and resequenced, or which has been
25 assembled from the overlapping sequences of more than one Incyte Clone using a computer program for fragment assembly, such as the GELVIEW™ Fragment Assembly system (GCG, Madison, WI). Some sequences have been both extended and assembled to produce the consensus sequence .

As used herein, the term "correlates with expression of a polynucleotide" indicates
30 that the detection of the presence of nucleic acids, the same or related to a nucleic acid sequence encoding HUPM, by northern analysis is indicative of the presence of nucleic acids encoding HUPM in a sample, and thereby correlates with expression of the transcript

from the polynucleotide encoding HUPM.

A "deletion," as the term is used herein, refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

5 The term "derivative," as used herein, refers to the chemical modification of HUPM, of a polynucleotide sequence encoding HUPM, or of a polynucleotide sequence complementary to a polynucleotide sequence encoding HUPM. Chemical modifications of a polynucleotide sequence can include, for example, replacement of hydrogen by an alkyl, acyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at
10 least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains a at least one biological or immunological function of the polypeptide from which it was derived.

The term "homology," as used herein, refers to a degree of complementarity.

15 There may be partial homology or complete homology. The word "identity" may substitute for the word "homology." A partially complementary sequence that at least partially inhibits an identical sequence from hybridizing to a target nucleic acid is referred to as "substantially homologous." The inhibition of hybridization of the completely complementary sequence to the target sequence may be examined using a hybridization
20 assay (Southern or northern blot, solution hybridization, and the like) under conditions of reduced stringency. A substantially homologous sequence or hybridization probe will compete for and inhibit the binding of a completely homologous sequence to the target sequence under conditions of reduced stringency. This is not to say that conditions of reduced stringency are such that non-specific binding is permitted, as reduced stringency
25 conditions require that the binding of two sequences to one another be a specific (i.e., a selective) interaction. The absence of non-specific binding may be tested by the use of a second target sequence which lacks even a partial degree of complementarity (e.g., less than about 30% homology or identity). In the absence of non-specific binding, the substantially homologous sequence or probe will not hybridize to the second non-
30 complementary target sequence.

The phrases "percent identity" or "% identity" refer to the percentage of sequence similarity found in a comparison of two or more amino acid or nucleic acid sequences.

Percent identity can be determined electronically, e.g., by using the MegAlign program (DNASTAR, Inc., Madison WI). This program can create alignments between two or more sequences according to different methods, e.g., the clustal method. (Higgins, D.G. and P. M. Sharp (1988) *Gene* 73:237-244.) The clustal algorithm groups sequences into
5 clusters by examining the distances between all pairs. The clusters are aligned pairwise and then in groups. The percentage similarity between two amino acid sequences, e.g., sequence A and sequence B, is calculated by dividing the length of sequence A, minus the number of gap residues in sequence A, minus the number of gap residues in sequence B, into the sum of the residue matches between sequence A and sequence B, times one
10 hundred. Gaps of low or of no homology between the two amino acid sequences are not included in determining percentage similarity. Percent identity between nucleic acid sequences can also be counted or calculated by other methods known in the art, such as the Jotun Hein method. (See, e.g., Hein, J. (1990) *Methods Enzymol.* 183:626-645.) Identity between sequences can also be determined by other methods known in the art, e.g., by
15 varying hybridization conditions.

“Human artificial chromosomes” (HACs), as described herein, are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size, and which contain all of the elements required for stable mitotic chromosome segregation and maintenance. (See, e.g., Harrington, J.J. et al. (1997) *Nat Genet.* 15:345-355.)

20 The term “humanized antibody,” as used herein, refers to antibody molecules in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

“Hybridization,” as the term is used herein, refers to any process by which a strand
25 of nucleic acid binds with a complementary strand through base pairing.

As used herein, the term “hybridization complex” as used herein, refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C_0t or R_0t analysis) or formed between one nucleic acid sequence present
30 in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" or "addition," as used herein, refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively, to the sequence found in the naturally occurring molecule.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

The term "microarray," as used herein, refers to an arrangement of distinct polynucleotides or oligonucleotides on a substrate, such as paper, nylon or any other type of membrane, filter, chip, glass slide, or any other suitable solid support.

The term "modulate," as it appears herein, refers to a change in the activity of HUPM. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of HUPM.

The phrases "nucleic acid" or "nucleic acid sequence," as used herein, refer to an oligonucleotide, nucleotide, polynucleotide, or any fragment thereof, to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material. In this context, "fragments" refers to those nucleic acid sequences which are greater than about 60 nucleotides in length, and most preferably are at least about 100 nucleotides, at least about 1000 nucleotides, or at least about 10,000 nucleotides in length.

The terms "operably associated" or "operably linked," as used herein, refer to functionally related nucleic acid sequences. A promoter is operably associated or operably linked with a coding sequence if the promoter controls the transcription of the encoded polypeptide. While operably associated or operably linked nucleic acid sequences can be contiguous and in reading frame, certain genetic elements, e.g., repressor genes, are not contiguously linked to the encoded polypeptide but still bind to operator sequences that control expression of the polypeptide.

The term "oligonucleotide," as used herein, refers to a nucleic acid sequence of at least about 6 nucleotides to 60 nucleotides, preferably about 15 to 30 nucleotides, and most preferably about 20 to 25 nucleotides, which can be used in PCR amplification or in

a hybridization assay or microarray. As used herein, the term "oligonucleotide" is substantially equivalent to the terms "amplimer," "primer," "oligomer," and "probe," as these terms are commonly defined in the art.

"Peptide nucleic acid" (PNA), as used herein, refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA and RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell. (See, e.g., Nielsen, P.E. et al. (1993) Anticancer Drug Des. 8:53-63.)

The term "sample," as used herein, is used in its broadest sense. A biological sample suspected of containing nucleic acids encoding HUPM, or fragments thereof, or HUPM itself, may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a solid support; a tissue; a tissue print; etc.

As used herein, the terms "specific binding" or "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, or an antagonist. The interaction is dependent upon the presence of a particular structure of the protein, the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide containing the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

As used herein, the term "stringent conditions" refers to conditions which permit hybridization between polynucleotide sequences and the claimed polynucleotide sequences. Suitably stringent conditions can be defined by, for example, the concentrations of salt or formamide in the prehybridization and hybridization solutions, or by the hybridization temperature, and are well known in the art. In particular, stringency can be increased by reducing the concentration of salt, increasing the concentration of formamide, or raising the hybridization temperature.

For example, hybridization under high stringency conditions could occur in about 50% formamide at about 37°C to 42°C. Hybridization could occur under reduced stringency conditions in about 35% to 25% formamide at about 30°C to 35°C. In

particular, hybridization could occur under high stringency conditions at 42°C in 50% formamide, 5X SSPE, 0.3% SDS, and 200 µg/ml sheared and denatured salmon sperm DNA. Hybridization could occur under reduced stringency conditions as described above, but in 35% formamide at a reduced temperature of 35°C. The temperature range

5 corresponding to a particular level of stringency can be further narrowed by calculating the purine to pyrimidine ratio of the nucleic acid of interest and adjusting the temperature accordingly. Variations on the above ranges and conditions are well known in the art.

The term "substantially purified," as used herein, refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or
10 separated, and are at least about 60% free, preferably about 75% free, and most preferably about 90% free from other components with which they are naturally associated.

A "substitution," as used herein, refers to the replacement of one or more amino acids or nucleotides by different amino acids or nucleotides, respectively.

"Transformation," as defined herein, describes a process by which exogenous DNA
15 enters and changes a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, viral infection,
20 electroporation, heat shock, lipofection, and particle bombardment. The term "transformed" cells includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, and transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

25 A "variant" of HUPM, as used herein, refers to an amino acid sequence that is altered by one or more amino acids. The variant may have "conservative" changes, wherein a substituted amino acid has similar structural or chemical properties (e.g., replacement of leucine with isoleucine). More rarely, a variant may have "nonconservative" changes (e.g., replacement of glycine with tryptophan). Analogous
30 minor variations may also include amino acid deletions or insertions, or both. Guidance in determining which amino acid residues may be substituted, inserted, or deleted without abolishing biological or immunological activity may be found using computer programs

well known in the art, for example, DNASTAR software.

THE INVENTION

The invention is based on the discovery of new human protease molecules
 5 (HUPM), the polynucleotides encoding HUPM, and the use of these compositions for the
 diagnosis, treatment, or prevention of cell proliferative and immune disorders. Table 1
 shows the sequence identification numbers, Incyte Clone identification number, and
 cDNA library for each of the human protease molecules disclosed herein.

10

15

Table 1

PROTEIN	NUCLEOTIDE	CLONE ID	LIBRARY
SEQ ID NO:1	SEQ ID NO:13	135360	BMARNOT02
SEQ ID NO:2	SEQ ID NO:14	447484	TYMNOT02
SEQ ID NO:3	SEQ ID NO:15	789927	PROSTUT03
SEQ ID NO:4	SEQ ID NO:16	877617	LUNGAST01
SEQ ID NO:5	SEQ ID NO:17	999322	KIDNTUT01
SEQ ID NO:6	SEQ ID NO:18	1337018	COLNNOT13
SEQ ID NO:7	SEQ ID NO:19	1798496	COLNNOT27
SEQ ID NO:8	SEQ ID NO:20	2082147	UTRSNOT08
SEQ ID NO:9	SEQ ID NO:21	2170967	ENDCNOT03
SEQ ID NO:10	SEQ ID NO:22	2484218	SMCANOT01
SEQ ID NO:11	SEQ ID NO:23	2680548	SINIUCT01
SEQ ID NO:12	SEQ ID NO:24	2957969	KIDNFET01

Nucleic acids encoding the HUPM-1 of the present invention were first identified

in Incyte Clone 135360 from the bone marrow cDNA library (BMARNOT02) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:13, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 135360 (BMARNOT02), 1440654 (THYRNOT03), 1985677
5 (LUNGAST01), 2016316 (ENDCNOT03), 2309369 (NGANNOT01), 3003105 (TYLMNOT06), and 3604791 (LUNGNOT30).

In one embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:1. HUPM-1 is 63 amino acids in length and, as shown in Figures 1A and 1B, has chemical and structural homology with rat proteasome
10 subunit, C8 (GI 203207). In particular, HUPM-1 and rat C8 share 54% identity. The fragment of SEQ ID NO:13 from about nucleotide 688 to about nucleotide 744 is useful for hybridization. Northern analysis shows the expression of this sequence in cardiovascular, male and female reproductive, and gastrointestinal cDNA libraries. Approximately 25% of these libraries are associated with neoplastic disorders and 33%
15 with inflammation and the immune response.

Nucleic acids encoding the HUPM-2 of the present invention were first identified in Incyte Clone 447484 from the T-lymphocyte cDNA library (TYLMNOT02) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:14, was derived from the following overlapping and/or extended nucleic acid
20 sequences: Incyte Clones 007562 (HMC1NOT01), 288369 (EOSIHET02), 447484 (TYLMNOT02), 1357876 (LUNGNOT09), 1688150 (PROSTUT10), 2506075 (CONUTUT01), 2748364 (LUNGTUT11), and shotgun sequences SAJA02963, SAJA00487, and SAJA00384.

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:2. HUPM-2 is 262 amino acids in length and has a
25 potential N-glycosylation site at N91, and potential phosphorylation sites for casein kinase II at S55, S63, S97, and T168, and for protein kinase C at S97, S186, and T246. A potential catalytic active site triad for cysteine proteases is found in amino acid residues C36, D176, and H177. The fragment of SEQ ID NO:14 from about nucleotide 2242 to
30 2292 encompasses the active site cysteine encoding region of the molecule and is useful for hybridization. Northern analysis shows the expression of this sequence in cardiovascular, male and female reproductive, and hematopoietic cDNA libraries.

Approximately 48% of these libraries are associated with neoplastic disorders and 24% with inflammation and the immune response.

Nucleic acids encoding the HUPM-3 of the present invention were first identified in Incyte Clone 789927 from the prostate tumor cDNA library (PROSTUT03) using a
5 computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:15, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 789927 (PROSTUT03), 1646976 (PROSTUT09), and 1979791 (LUNGTUT03).

In another embodiment, the invention encompasses a polypeptide comprising the
10 amino acid sequence of SEQ ID NO:3. HUPM-3 is 314 amino acids in length and has a potential signal peptide sequence between amino acid residues M1 and R19. Potential N-glycosylation sites are found at residues N167, N200, and N273, and potential phosphorylation sites are found for casein kinase II at T86, S134, S161, T190, and S291, and for protein kinase C at T39, S58, S73, S127, and S212. Sequences containing
15 potential active site histidine and serine residues, characteristic of serine proteases, are found at LTAAH82 and GDS238GGP in HUPM-3. The fragment of SEQ ID NO:15 between about nucleotide 271 to about nucleotide 330 which encompasses the active site histidine is useful for hybridization. Northern analysis shows the expression of this sequence in cardiovascular, hematopoietic, and male reproductive cDNA libraries.
20 Approximately 86% of these libraries are associated with neoplastic disorders.

Nucleic acids encoding the HUPM-4 of the present invention were first identified in Incyte Clone 877617 from the lung cDNA library (LUNGAST01) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:16, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte
25 Clones 372314 (LUNGNOT02), 698335 (SYNORAT03), 692718 (LUNGTUT02), 877617 (LUNGAST01), and 1399470 (BRAITUT08).

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:4. HUPM-4 is 420 amino acids in length and has a potential signal peptide sequence extending from residues M1 to P21. Potential N-
30 glycosylation sites are found at residues N90, N133, and N336. Potential phosphorylation sites are found for casein kinase II at S60 and T338, and for protein kinase C at S106, T143, T346, and S393. Two potential leucine zipper patterns are found beginning at

L309 and L316, and a potential cell attachment site is found in the sequence R387GD.

Two potential active site aspartate residues, characteristic of aspartic proteases, are found at residues D96 and D283. The fragment of SEQ ID NO:16 from about nucleotide 1609 to about nucleotide 1692, encompassing a leucine zipper domain, is useful for hybridization.

- 5 Northern analysis shows the expression of this sequence in cardiovascular, hematopoietic, and male and female reproductive cDNA libraries. Approximately 56% of these libraries are associated with neoplastic disorders, 18% with inflammation and the immune response, and 18% with trauma.

Nucleic acids encoding the HUPM-5 of the present invention were first identified
10 in Incyte Clone 999322 from the kidney tumor cDNA library (KIDNTUT01) using a computer search for amino acid sequence alignments, and a consensus sequence, SEQ ID NO:17, was derived from this clone.

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:5. HUPM-5 is 200 amino acids in length and has a
15 potential N-glycosylation site at N121, and potential phosphorylation sites for cAMP- and cGMP-dependent protein kinase at S35, for casein kinase II at S150 and T158, and for protein kinase C at T180. A potential active site serine for serine protease is found in the sequence GDS112GGP. The fragment of SEQ ID NO:17 from about nucleotide 775 to about nucleotide 838 from the active site serine domain is useful for hybridization.

- 20 Northern analysis shows the expression of this sequence exclusively in kidney tumor (KIDNTUT01).

Nucleic acids encoding the HUPM-6 of the present invention were first identified in Incyte Clone 1337018 from the colon cDNA library (COLNNOT13) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:18, was
25 derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 1271725 (TESTTUT02), 1337018 (COLNNOT13), 586982 and 588598 (UTRSNOT01).

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:6. HUPM-6 is 435 amino acids in length and has
30 potential N-glycosylation sites at residues N128 and N176, potential phosphorylation sites for cAMP- and cGMP-dependent protein kinase T249, for casein kinase II at S93 and S231, for protein kinase C at T26, S144, T148, S197, T200, S260, T303, S351, and T365,

and for tyrosine kinase at Y59 and Y360. Sequences containing potential active site histidine and serine residues for serine proteases are found at LTAAH243C and GDS385GGP, respectively. The fragment of SEQ ID NO:18 from about nucleotide 900 to about nucleotide 949 encompassing the active site histidine residue is useful for

5 hybridization. Northern analysis shows the expression of this sequence in gastrointestinal and male and female reproductive cDNA libraries. Approximately 65% of these libraries are associated with neoplastic disorders and 22% with the immune response.

Nucleic acids encoding the HUPM-7 of the present invention were first identified in Incyte Clone 1798496 from the colon cDNA library (COLNNOT27) using a computer

10 search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:19, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 819896 (KERANOT02), 1798496 (COLNNOT27), and shotgun sequence SAGA00119.

In another embodiment, the invention encompasses a polypeptide comprising the

15 amino acid sequence of SEQ ID NO:7. HUPM-7 is 260 amino acids in length and has a potential signal peptide sequence extending from residues M1 to A28. Potential N-myristoylation sites are found in the vicinity of the signal peptide cleavage site at G19, G20, and G35. A potential N-glycosylation site is found at N110, and potential phosphorylation sites are found for casein kinase II at S112, S140, and S162, for protein

20 kinase C at T80, S162, S201, and S236, and for tyrosine kinase at Y188. A potential glycosaminoglycan attachment site is found at S155, and sequences containing potential active site histidine and serine residues for serine proteases are found at LTAAH73C and GDS212GGP, respectively. The fragment of SEQ ID NO:19 from about nucleotide 517 to about nucleotide 574, located between the active site histidine and serine residues, is

25 useful for hybridization. Northern analysis shows the expression of this sequence in female reproductive, neural, lung, and colon cDNA libraries. Approximately 83% of these libraries are associated with neoplastic disorders.

Nucleic acids encoding the HUPM-8 of the present invention were first identified in Incyte Clone 2082147 from the uterine tissue cDNA library (UTRSNOT08) using a

30 computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:20, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 586776 (UTRSNOT01), 1719194 (BLADNOT06), 2082147 and

2082170 (UTRSNOT08), 3359814 (PROSTUT16), and shotgun sequences SAGA01368 and SAGA01895.

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:8. HUPM-8 is 175 amino acids in length and has a
5 potential signal peptide sequence extending between residues M1 and L30, potential phosphorylation sites for casein kinase II at T28, and for protein kinase C at S81. A potential cell attachment site sequence is found at R73DG, and a potential signal peptidase signature sequence containing an active site serine residue is found in the sequence GDHHGHS128FD. The fragment of SEQ ID NO:20 from about nucleotide 757 to about
10 nucleotide 789 from the catalytic active site is useful for hybridization. Northern analysis shows the expression of this sequence in fetal, gastrointestinal, male and female reproductive, and neuronal cDNA libraries. Approximately 38% of these libraries are associated with neoplastic disorders, 24% with the immune response, and 14% with fetal development.

15 Nucleic acids encoding the HUPM-9 of the present invention were first identified in Incyte Clone 2170967 from the endothelial cell cDNA library (ENDCNOT03) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:21, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 1667462 (BMARNOT03), 1830465 (THP1AZT01), 1888989
20 (BLADTUT07), 1928627 (BRSTNOT02), 2170967 (ENDCNOT03), 3125590 (LUNGTUT12), and 3456567 (293TF1T01).

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:9. HUPM-9 is 519 amino acids in length and has a potential aminopeptidase signature sequence at N362TDAEGRL in which D364 and E366
25 represent the zinc binding ligands at the active site. HUPM-9 also has two potential N-glycosylation sites at N72 and N410, and potential phosphorylation sites for casein kinase II at S28, S54, S138, S228, S238, T363, T487, and T506, and for protein kinase C at S174, S227, S292, S340, T487, and T500. The fragment of SEQ ID NO:21 from about nucleotide 688 to about nucleotide 747 is useful for hybridization. Northern analysis
30 shows the expression of this sequence in cardiovascular, male and female reproductive, hematopoietic, and nervous system cDNA libraries. Approximately 46% of these libraries are associated with neoplastic disorders and 31% with the immune response.

Nucleic acids encoding the HUPM-10 of the present invention were first identified in Incyte Clone 2484218 from the aortic smooth muscle cell cDNA library (SMCANOT01) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:22, was derived from the following overlapping and/or
5 extended nucleic acid sequences: Incyte Clones 1351043 (LATRTUT02), 1381980 (BRAITUT08), 1432027 (BEPINON01), 1457881 (COLNFET02), and 2484218 (SMCANOT01).

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:10. HUPM-10 is 327 amino acids in length and has
10 three potential N-glycosylation sites at N12, N50, and N214, and potential phosphorylation sites for casein kinase II at S18, T93, T107, S166, S170, and T216, for protein kinase C at T272, and for tyrosine kinase at Y104. HUPM-10 has chemical and structural homology with human proteasome subunit p40 (GI 971270). In particular, HUPM-10 and human p40 share 23% homology. The fragment of SEQ ID NO:22 from
15 about nucleotide 136 to about nucleotide 211 is useful for hybridization. Northern analysis shows the expression of this sequence in cardiovascular, male and female reproductive, nervous system, and hematopoietic cDNA libraries. Approximately 40% of these libraries are associated with neoplastic disorders, 24% with the immune response, and 22% with fetal development.

20 Nucleic acids encoding the HUPM-11 of the present invention were first identified in Incyte Clone 2680548 from the ileum tissue cDNA library (SINIUCT01) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:23, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 725100 (SYNOOAT01), 779975 (MYOMNOT01), 1528274
25 (UCMCL5T01), 1658964 (URETTUT01), 1781933 (PGANNON02), 2618786 (GBLANOT01), and 2680548 (SINIUCT01).

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:11. HUPM-11 is 458 amino acids in length and has two sequences containing potential active site histidine and serine residues for serine
30 proteases at VTNAH198V and GNS306GGP, respectively. Two potential N-glycosylation sites are found at N181 and N349, and potential phosphorylation sites are found for cAMP- and cGMP-dependent protein kinase at S350, for casein kinase II at

T221, T290, and S383, and for protein kinase C at S13, S142, T231, T322, S335, and S357. The fragment of SEQ ID NO:23 from about nucleotide 694 to about nucleotide 756, located between the potential histidine and serine active site residues, is useful for hybridization. Northern analysis shows the expression of this sequence in gastrointestinal, male and female reproductive, and nervous system cDNA libraries. Approximately 43% of these libraries are associated with neoplastic disorders and 25% with the immune response.

Nucleic acids encoding the HUPM-12 of the present invention were first identified in Incyte Clone 2957969 from the fetal kidney cDNA library (KIDNFET01) using a computer search for amino acid sequence alignments. A consensus sequence, SEQ ID NO:24, was derived from the following overlapping and/or extended nucleic acid sequences: Incyte Clones 875973 (LUNGAST01), 978220 (BRSTNOT02), 1362955 (LUNGNOT12), 1511581 (LUNGNOT14), 2354566 (LUNGNOT20), 2957969 (KIDNFET01), and 3676880 (PLACNOT07).

In another embodiment, the invention encompasses a polypeptide comprising the amino acid sequence of SEQ ID NO:12. HUPM-12 is 532 amino acids in length and has three potential N-glycosylation sites at N182, N329, and N348, potential phosphorylation sites for casein kinase II at S20, T205, T331, T350, and S441, and for protein kinase C at T144, S150, S279, S341, T388, and S526. A potential aminopeptidase signature sequence is found at N349TDAEGRL in which D351 and E353 represent the zinc binding ligands at the active site. A potential ATP/GTP-binding site (P-loop) is also found in the sequence G277LSIKGKT. The fragment of SEQ ID NO:24 from about nucleotide 709 to about nucleotide 781 is useful for hybridization. Northern analysis shows the expression of this sequence in cardiovascular, male and female reproductive, and nervous system cDNA libraries. Approximately 55% of these libraries are associated with neoplastic disorders, 12% with the immune response, and 14% with fetal tissues and proliferative cell lines.

The invention also encompasses HUPM variants. A preferred HUPM variant is one which has at least about 80%, more preferably at least about 90%, and most preferably at least about 95% amino acid sequence identity to the HUPM amino acid sequence, and which contains at least one functional or structural characteristic of HUPM.

The invention also encompasses polynucleotides which encode HUPM. In a particular embodiment, the invention encompasses a polynucleotide consisting of a

nucleic acid sequence selected from the group consisting of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24.

The invention also encompasses a variant of a polynucleotide sequence encoding
5 HUPM. In particular, such a variant polynucleotide sequence will have at least about 80%, more preferably at least about 90%, and most preferably at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding HUPM. A particular aspect of the invention encompasses a variant of a polynucleotide sequence selected from the group consisting of SEQ ID NO:13 SEQ ID NO:14, SEQ ID NO:15,
10 SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24 which has at least about 80%, more preferably at least about 90%, and most preferably at least about 95% polynucleotide sequence identity to a polynucleotide sequence selected from the group consisting of SEQ ID NO:13 SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID
15 NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of HUPM.

It will be appreciated by those skilled in the art that as a result of the degeneracy of
20 the genetic code, a multitude of polynucleotide sequences encoding HUPM, some bearing minimal homology to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard
25 triplet genetic code as applied to the polynucleotide sequence of naturally occurring HUPM, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode HUPM and its variants are preferably capable of hybridizing to the nucleotide sequence of the naturally occurring HUPM under appropriately selected conditions of stringency, it may be advantageous to
30 produce nucleotide sequences encoding HUPM or its derivatives possessing a substantially different codon usage. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the

frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding HUPM and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced
5 from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode HUPM and HUPM derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents that are well known in the art.
10 Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding HUPM or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ
15 ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, or SEQ ID NO:24, or fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) *Methods Enzymol.* 152:399-407; and Kimmel, A.R. (1987) *Methods Enzymol.* 152:507-511.)

Methods for DNA sequencing are well known and generally available in the art
20 and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, Sequenase® (US Biochemical Corp., Cleveland, OH), Taq polymerase (Perkin Elmer), thermostable T7 polymerase (Amersham, Chicago, IL), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE Amplification System (GIBCO/BRL,
25 Gaithersburg, MD). Preferably, the process is automated with machines such as the Hamilton Micro Lab 2200 (Hamilton, Reno, NV), Peltier Thermal Cycler (PTC200; MJ Research, Watertown, MA) and the ABI Catalyst and 373 and 377 DNA Sequencers (Perkin Elmer).

The nucleic acid sequences encoding HUPM may be extended utilizing a partial
30 nucleotide sequence and employing various methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal primers to retrieve unknown

sequence adjacent to a known locus. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) In particular, genomic DNA is first amplified in the presence of a primer complementary to a linker sequence within the vector and a primer specific to a region of the nucleotide sequence. The amplified sequences are then subjected to a second round of

5 PCR with the same linker primer and another specific primer internal to the first one. Products of each round of PCR are transcribed with an appropriate RNA polymerase and sequenced using reverse transcriptase.

Inverse PCR may also be used to amplify or extend sequences using divergent primers based on a known region. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) The primers may be designed using commercially available software such as OLIGO 4.06 Primer Analysis software (National Biosciences Inc., Plymouth, MN) or another appropriate program to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to 72°C. The method uses several restriction enzymes to generate a suitable

15 fragment in the known region of a gene. The fragment is then circularized by intramolecular ligation and used as a PCR template.

Another method which may be used is capture PCR, which involves PCR amplification of DNA fragments adjacent to a known sequence in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods

20 Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to place an engineered double-stranded sequence into an unknown fragment of the DNA molecule before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060.) Additionally, one may use PCR, nested primers, and

25 PromoterFinder™ libraries to walk genomic DNA (Clontech, Palo Alto, CA). This process avoids the need to screen libraries and is useful in finding intron/exon junctions.

When screening for full-length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. Also, random-primed libraries are preferable in that they will include more sequences which contain the 5' regions of genes. Use of a

30 randomly primed library may be especially preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different fluorescent dyes (one for each nucleotide) which are laser
5 activated, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., Genotyper™ and Sequence Navigator™, Perkin Elmer), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for the sequencing of small
10 pieces of DNA which might be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode HUPM may be used in recombinant DNA molecules to direct expression of HUPM, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which
15 encode substantially the same or a functionally equivalent amino acid sequence may be produced, and these sequences may be used to clone and express HUPM.

As will be understood by those of skill in the art, it may be advantageous to produce HUPM-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can
20 be selected to increase the rate of protein expression or to produce an RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter HUPM-encoding sequences for a
25 variety of reasons including, but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation patterns, change codon
30 preference, produce splice variants, introduce mutations, and so forth.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding HUPM may be ligated to a heterologous sequence to encode a

fusion protein. For example, to screen peptide libraries for inhibitors of HUPM activity, it may be useful to encode a chimeric HUPM protein that can be recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the HUPM encoding sequence and the heterologous protein
5 sequence, so that HUPM may be cleaved and purified away from the heterologous moiety.

In another embodiment, sequences encoding HUPM may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucl. Acids Res. Symp. Ser. 215-223, and Horn, T. et al. (1980) Nucl. Acids Res. Symp. Ser. 225-232.) Alternatively, the protein itself may be produced using chemical
10 methods to synthesize the amino acid sequence of HUPM, or a fragment thereof. For example, peptide synthesis can be performed using various solid-phase techniques. (See, e.g., Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A Peptide Synthesizer (Perkin Elmer). Additionally, the amino acid sequence of HUPM, or any part thereof, may be altered during direct synthesis
15 and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid
20 analysis or by sequencing. (See, e.g., Creighton, T. (1983) Proteins, Structures and Molecular Properties, WH Freeman and Co., New York, NY.)

In order to express a biologically active HUPM, the nucleotide sequences encoding HUPM or derivatives thereof may be inserted into appropriate expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the
25 inserted coding sequence.

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding HUPM and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook,
30 J. et al. (1989) Molecular Cloning. A Laboratory Manual, Cold Spring Harbor Press, Plainview, NY, ch. 4, 8, and 16-17; and Ausubel, F.M. et al. (1995, and periodic supplements) Current Protocols in Molecular Biology, John Wiley & Sons, New York,

NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding HUPM. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression
5 vectors; yeast transformed with yeast expression vectors; insect cell systems infected with virus expression vectors (e.g., baculovirus); plant cell systems transformed with virus expression vectors (e.g., cauliflower mosaic virus (CaMV) or tobacco mosaic virus (TMV)) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems.

10 The invention is not limited by the host cell employed.

The "control elements" or "regulatory sequences" are those non-translated regions, e.g., enhancers, promoters, and 5' and 3' untranslated regions, of the vector and polynucleotide sequences encoding HUPM which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and
15 specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters, e.g., hybrid lacZ promoter of the Bluescript® phagemid (Stratagene, La Jolla, CA) or pSport1™ plasmid (GIBCO/BRL), may be used. The baculovirus polyhedrin promoter may be used
20 in insect cells. Promoters or enhancers derived from the genomes of plant cells (e.g., heat shock, RUBISCO, and storage protein genes) or from plant viruses (e.g., viral promoters or leader sequences) may be cloned into the vector. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are preferable. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding
25 HUPM, vectors based on SV40 or EBV may be used with an appropriate selectable marker.

In bacterial systems, a number of expression vectors may be selected depending upon the use intended for HUPM. For example, when large quantities of HUPM are needed for the induction of antibodies, vectors which direct high level expression of fusion
30 proteins that are readily purified may be used. Such vectors include, but are not limited to, multifunctional E. coli cloning and expression vectors such as Bluescript® (Stratagene), in which the sequence encoding HUPM may be ligated into the vector in frame with

sequences for the amino-terminal Met and the subsequent 7 residues of β -galactosidase so that a hybrid protein is produced, and pSPORT vectors. (Gibco/BRL, Gaithersburg, MD.) pGEX vectors (Pharmacia Biotech, Uppsala, Sweden) may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such

5 fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

10 In the yeast Saccharomyces cerevisiae, a number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH, may be used. (See, e.g., Ausubel, supra; and Grant et al. (1987) Methods Enzymol. 153:516-544.)

In cases where plant expression vectors are used, the expression of sequences encoding HUPM may be driven by any of a number of promoters. For example, viral

15 promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV. (Takamatsu, N. (1987) EMBO J. 6:307-311.) Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews. (See, e.g., Hobbs, S. or Murry, L.E. in McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, NY; pp. 191-196.)

20

25 An insect system may also be used to express HUPM. For example, in one such system, Autographa californica nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in Spodoptera frugiperda cells or in Trichoplusia larvae. The sequences encoding HUPM may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful

30 insertion of sequences encoding HUPM will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses may then be used to infect, for example, S. frugiperda cells or Trichoplusia larvae in which HUPM may

be expressed. (See, e.g., Engelhard, E.K. et al. (1994) Proc. Nat. Acad. Sci. 91:3224-3227.)

In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding
5 HUPM may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing HUPM in infected host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. 81:3655-3659.) In addition, transcription enhancers, such as the Rous
10 sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained and expressed in a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes,
15 polycationic amino polymers, or vesicles) for therapeutic purposes.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding HUPM. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding HUPM and its initiation codon and upstream sequences are inserted into the appropriate expression vector, no additional
20 transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both
25 natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such
30 modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct

insertion, folding, and/or function. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38), are available from the American Type Culture Collection (ATCC, Bethesda, MD) and may be chosen to ensure the correct modification
5 and processing of the foreign protein.

For long term, high yield production of recombinant proteins, stable expression is preferred. For example, cell lines capable of stably expressing HUPM can be transformed using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector.
10 Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to
15 the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase genes and adenine phosphoribosyltransferase genes, which can be employed in *tk* or *ap^r* cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; and Lowy, I. et al.
20 (1980) Cell 22:817-823) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *npt* confers resistance to the aminoglycosides neomycin and G-418; and *als* or *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. 77:3567-3570; Colbere-Garapin, F. et al
25 (1981) J. Mol. Biol. 150:1-14; and Murry, supra.) Additional selectable genes have been described, e.g., *trpB*, which allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histinol in place of histidine. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. 85:8047-8051.) Recently, the use of visible
30 markers, such as anthocyanins, green fluorescent proteins, β glucuronidase and its substrate GUS, luciferase and its substrate luciferin, has increased. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A.

et al. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding HUPM is inserted within a marker gene sequence,
5 transformed cells containing sequences encoding HUPM can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding HUPM under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

10 Alternatively, host cells which contain the nucleic acid sequence encoding HUPM and express HUPM may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid
15 or protein sequences.

The presence of polynucleotide sequences encoding HUPM can be detected by DNA-DNA or DNA-RNA hybridization or amplification using probes or fragments or fragments of polynucleotides encoding HUPM. Nucleic acid amplification based assays involve the use of oligonucleotides or oligomers based on the sequences encoding HUPM
20 to detect transformants containing DNA or RNA encoding HUPM.

A variety of protocols for detecting and measuring the expression of HUPM, using either polyclonal or monoclonal antibodies specific for the protein, are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site,
25 monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on HUPM is preferred, but a competitive binding assay may be employed. These and other assays are well described in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St Paul, MN, Section IV; and Maddox, D.E. et al. (1983) J. Exp. Med. 158:1211-1216).

30 A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to

polynucleotides encoding HUPM include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding HUPM, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be
5 used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Pharmacia & Upjohn (Kalamazoo, MI), Promega (Madison, WI), and U.S. Biochemical Corp. (Cleveland, OH). Suitable reporter molecules or labels which may be used for ease of detection include
10 radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding HUPM may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or contained
15 intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode HUPM may be designed to contain signal sequences which direct secretion of HUPM through a prokaryotic or eukaryotic cell membrane. Other constructions may be used to join sequences encoding HUPM to nucleotide sequences encoding a polypeptide domain
20 which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, WA). The
25 inclusion of cleavable linker sequences, such as those specific for Factor XA or enterokinase (Invitrogen, San Diego, CA), between the purification domain and the HUPM encoding sequence may be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing HUPM and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site.
30 The histidine residues facilitate purification on immobilized metal ion affinity chromatography. (IMAC) (See, e.g., Porath, J. et al. (1992) Prot. Exp. Purif. 3: 263-281.) The enterokinase cleavage site provides a means for purifying HUPM from the fusion

protein. (See, e.g., Kroll, D.J. et al. (1993) DNA Cell Biol. 12:441-453.)

Fragments of HUPM may be produced not only by recombinant production, but also by direct peptide synthesis using solid-phase techniques. (See, e.g., Creighton, T.E. (1984) Protein: Structures and Molecular Properties, pp. 55-60, W.H. Freeman and Co., New York, NY.) Protein synthesis may be performed by manual techniques or by automation. Automated synthesis may be achieved, for example, using the Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Various fragments of HUPM may be synthesized separately and then combined to produce the full length molecule.

10 THERAPEUTICS

Chemical and structural homology exists among the human protease molecules of the invention. In addition, HUPM is expressed in proliferating cell types associated with cancer, and the immune response. Therefore, HUPM appears to play a role in cell proliferative disorders and immune disorders. Therefore, in cell proliferative or immune disorders where HUPM is being expressed or is promoting cell proliferation it is desirable to decrease the expression of HUPM. In cell proliferative or immune disorders where expression of HUPM is decreased, it is desirable to provide the protein or increase expression.

Therefore, in one embodiment, an antagonist of HUPM may be administered to a subject to treat or prevent a cell proliferative disorder associated with increased expression or activity of HUPM. Such a disorder may include, but is not limited to, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. In one aspect, an antibody which specifically binds HUPM may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissue which express HUPM.

In another additional embodiment, a vector expressing the complement of the

polynucleotide encoding HUPM may be administered to a subject to treat or prevent a cell proliferative disorder including, but not limited to, those described above.

In another embodiment, an antagonist of HUPM may be administered to a subject to treat or prevent an immune disorder associated with increased expression or activity of HUPM. Such a disorder may include, but is not limited to AIDS, Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, lupus erythematosus, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, ulcerative colitis, Werner syndrome, and complications of cancer, hemodialysis, and extracorporeal circulation; viral, bacterial, fungal, parasitic, protozoal, and helminthic infections; and trauma.

In still another embodiment, a vector expressing the complement of the polynucleotide encoding HUPM may be administered to a subject to treat or prevent an immune disorder including, but not limited to, those described above.

In another embodiment, HUPM or a fragment or derivative thereof may be administered to a subject to treat or prevent a cell proliferative disorder associated with decreased expression or activity of HUPM. Such disorders can include, but are not limited to, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing HUPM or a fragment or

derivative thereof may be administered to a subject to treat or prevent a cell proliferative disorder including, but not limited to, those described above.

In a further embodiment, a pharmaceutical composition comprising a substantially purified HUPM in conjunction with a suitable pharmaceutical carrier may be administered
5 to a subject to treat or prevent a cell proliferative disorder including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of HUPM may be administered to a subject to treat or prevent a cell proliferative disorder including, but not limited to, those listed above.

10 In another embodiment, HUPM or a fragment or derivative thereof may be administered to a subject to treat or prevent an immune disorder associated with decreased expression or activity of HUPM. Such disorders can include, but are not limited to, AIDS, Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia,
15 autoimmune thyroiditis, bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, lupus erythematosus, multiple sclerosis, myasthenia gravis, myocardial or pericardial
20 inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, ulcerative colitis, Werner syndrome, and complications of cancer, hemodialysis, and extracorporeal circulation; viral, bacterial, fungal, parasitic, protozoal, and helminthic infections; and trauma.

25 In another embodiment, a vector capable of expressing HUPM or a fragment or derivative thereof may be administered to a subject to treat or prevent an immune disorder including, but not limited to, those described above.

In a further embodiment, a pharmaceutical composition comprising a substantially purified HUPM in conjunction with a suitable pharmaceutical carrier may be administered
30 to a subject to treat or prevent an immune disorder including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of HUPM

may be administered to a subject to treat or prevent an immune disorder including, but not limited to, those listed above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination
5 with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower
10 dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of HUPM may be produced using methods which are generally known in the art. In particular, purified HUPM may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind HUPM. Antibodies to HUPM may also be generated using methods that are well known in the art.
15 Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are especially preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice,
20 humans, and others may be immunized by injection with HUPM or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil
25 emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to HUPM have an amino acid sequence consisting of at least about 5 amino acids, and, more preferably, of at least about 10 amino acids. It is also preferable that
30 these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein and contain the entire amino acid sequence of a small, naturally occurring molecule. Short stretches of HUPM amino acids may be fused with

those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to HUPM may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture.

5 These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) *Nature* 256:495-497; Kozbor, D. et al. (1985) *J. Immunol. Methods* 81:31-42; Cote, R.J. et al. (1983) *Proc. Natl. Acad. Sci.* 80:2026-2030; and Cole, S.P. et al. (1984) *Mol. Cell Biol.* 62:109-120.)

10 In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) *Proc. Natl. Acad. Sci.* 81:6851-6855; Neuberger, M.S. et al. (1984) *Nature* 312:604-608; and Takeda, S. et al. (1985) *Nature* 314:452-454.)

15 Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce HUPM-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton D.R. (1991) *Proc. Natl. Acad. Sci.* 88:10134-10137.)

20 Antibodies may also be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) *Proc. Natl. Acad. Sci.* 86: 3833-3837; and Winter, G. et al. (1991) *Nature* 349:293-299.)

Antibody fragments which contain specific binding sites for HUPM may also be
25 generated. For example, such fragments include, but are not limited to, F(ab')₂ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) *Science*
30 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric

assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between HUPM and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering HUPM epitopes is preferred, but a competitive binding assay may also be employed. (Maddox, supra.)

In another embodiment of the invention, the polynucleotides encoding HUPM, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, the complement of the polynucleotide encoding HUPM may be used in situations in which it would be desirable to block the transcription of the mRNA. In particular, cells may be transformed with sequences complementary to polynucleotides encoding HUPM. Thus, complementary molecules or fragments may be used to modulate HUPM activity, or to achieve regulation of gene function. Such technology is now well known in the art, and sense or antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding HUPM.

Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. Methods which are well known to those skilled in the art can be used to construct vectors which will express nucleic acid sequences complementary to the polynucleotides of the gene encoding HUPM. (See, e.g., Sambrook, supra; and Ausubel, supra.)

Genes encoding HUPM can be turned off by transforming a cell or tissue with expression vectors which express high levels of a polynucleotide, or fragment thereof, encoding HUPM. Such constructs may be used to introduce untranslatable sense or antisense sequences into a cell. Even in the absence of integration into the DNA, such vectors may continue to transcribe RNA molecules until they are disabled by endogenous nucleases. Transient expression may last for a month or more with a non-replicating vector, and may last even longer if appropriate replication elements are part of the vector system.

As mentioned above, modifications of gene expression can be obtained by designing complementary sequences or antisense molecules (DNA, RNA, or PNA) to the control, 5', or regulatory regions of the gene encoding HUPM. Oligonucleotides derived

from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, are preferred. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing Co., Mt. Kisco, NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

10 Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences
15 encoding HUPM.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene
20 containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be
25 prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding HUPM. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA
30 polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is
5 inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

Many methods for introducing vectors into cells or tissues are available and
10 equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nature Biotechnology 15:462-466.)

15 Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

An additional embodiment of the invention relates to the administration of a pharmaceutical or sterile composition, in conjunction with a pharmaceutically acceptable
20 carrier, for any of the therapeutic effects discussed above. Such pharmaceutical compositions may consist of HUPM, antibodies to HUPM, and mimetics, agonists, antagonists, or inhibitors of HUPM. The compositions may be administered alone or in combination with at least one other agent, such as a stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier including, but not limited
25 to, saline, buffered saline, dextrose, and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs, or hormones.

The pharmaceutical compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous,
30 intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically-acceptable carriers comprising excipients and auxiliaries

which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing Co., Easton, PA).

5 Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for ingestion by the patient.

10 Pharmaceutical preparations for oral use can be obtained through combining active compounds with solid excipient and processing the resultant mixture of granules (optionally, after grinding) to obtain tablets or dragee cores. Suitable auxiliaries can be added, if desired. Suitable excipients include carbohydrate or protein fillers, such as sugars, including lactose, sucrose, mannitol, and sorbitol; starch from corn, wheat, rice,
15 potato, or other plants; cellulose, such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethylcellulose; gums, including arabic and tragacanth; and proteins, such as gelatin and collagen. If desired, disintegrating or solubilizing agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, and alginic acid or a salt thereof, such as sodium alginate.

20 Dragee cores may be used in conjunction with suitable coatings, such as concentrated sugar solutions, which may also contain gum arabic, talc, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for product identification or to characterize the
25 quantity of active compound, i.e., dosage.

 Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating, such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with fillers or binders, such as lactose or starches, lubricants, such as talc or magnesium stearate, and,
30 optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid, or liquid polyethylene glycol with or without stabilizers.

Pharmaceutical formulations suitable for parenteral administration may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks's solution, Ringer's solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils, such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate, triglycerides, or liposomes. Non-lipid polycationic amino polymers may also be used for delivery. Optionally, the suspension may also contain suitable stabilizers or agents to increase the solubility of the compounds and allow for the preparation of highly concentrated solutions.

For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

The pharmaceutical compositions of the present invention may be manufactured in a manner that is known in the art, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping, or lyophilizing processes.

The pharmaceutical composition may be provided as a salt and can be formed with many acids, including but not limited to, hydrochloric, sulfuric, acetic, lactic, tartaric, malic, and succinic acid. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free base forms. In other cases, the preferred preparation may be a lyophilized powder which may contain any or all of the following: 1 mM to 50 mM histidine, 0.1% to 2% sucrose, and 2% to 7% mannitol, at a pH range of 4.5 to 5.5, that is combined with buffer prior to use.

After pharmaceutical compositions have been prepared, they can be placed in an appropriate container and labeled for treatment of an indicated condition. For administration of HUPM, such labeling would include amount, frequency, and method of administration.

Pharmaceutical compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those

skilled in the art.

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells or in animal models such as mice, rats, rabbits, dogs, or pigs. An animal model may also be used to determine the appropriate
5 concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example HUPM or fragments thereof, antibodies of HUPM, and agonists, antagonists or inhibitors of HUPM, which ameliorates the symptoms or condition. Therapeutic efficacy
10 and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED50 (the dose therapeutically effective in 50% of the population) or LD50 (the dose lethal to 50% of the population) statistics. The dose ratio of therapeutic to toxic effects is the therapeutic index, and it can be expressed as the ED50/LD50 ratio. Pharmaceutical compositions which exhibit large
15 therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED50 with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

20 The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration,
25 drug combination(s), reaction sensitivities, and response to therapy. Long-acting pharmaceutical compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1 μg to 100,000 μg , up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to
30 particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of

polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

5 In another embodiment, antibodies which specifically bind HUPM may be used for the diagnosis of disorders characterized by expression of HUPM, or in assays to monitor patients being treated with HUPM or agonists, antagonists, or inhibitors of HUPM. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for HUPM include methods which
10 utilize the antibody and a label to detect HUPM in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

15 A variety of protocols for measuring HUPM, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of HUPM expression. Normal or standard values for HUPM expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, preferably human, with antibody to HUPM under conditions suitable for complex formation. The
20 amount of standard complex formation may be quantitated by various methods, preferably by photometric means. Quantities of HUPM expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding HUPM may
25 be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantitate gene expression in biopsied tissues in which expression of HUPM may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of HUPM, and to monitor
30 regulation of HUPM levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding HUPM or closely

related molecules may be used to identify nucleic acid sequences which encode HUPM. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification (maximal, high, intermediate, or low), will
5 determine whether the probe identifies only naturally occurring sequences encoding HUPM, alleles, or related sequences.

Probes may also be used for the detection of related sequences, and should preferably contain at least 50% identity to the nucleotides from any of the HUPM encoding sequences. The hybridization probes of the subject invention may be DNA or
10 RNA and may be derived from the sequence of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, or SEQ ID NO:24 or from genomic sequences including promoters, enhancers, and introns of the HUPM gene.

Means for producing specific hybridization probes for DNAs encoding HUPM
15 include the cloning of polynucleotide sequences encoding HUPM or HUPM derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by
20 radionuclides such as ^{32}P or ^{35}S , or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding HUPM may be used for the diagnosis of a disorder associated with expression of HUPM. Examples of such a disorder include, but are not limited to, cell proliferative disorders such as arteriosclerosis, atherosclerosis,
25 bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia,
30 gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; and immune disorders such as AIDS, Addison's disease, adult respiratory distress syndrome,

allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, lupus erythematosus, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, ulcerative colitis, Werner syndrome, and complications of cancer, hemodialysis, and extracorporeal circulation; viral, bacterial, fungal, parasitic, protozoal, and helminthic infections; and trauma. The polynucleotide sequences encoding HUPM may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and ELISA assays; and in microarrays utilizing fluids or tissues from patients to detect altered HUPM expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding HUPM may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding HUPM may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantitated and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding HUPM in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of HUPM, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding HUPM, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an

experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

5 Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

10 With respect to cancer, the presence of a relatively high amount of transcript in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby
15 preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding HUPM may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding HUPM, or a fragment of a polynucleotide
20 complementary to the polynucleotide encoding HUPM, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantitation of closely related DNA or RNA sequences.

Methods which may also be used to quantitate the expression of HUPM include
25 radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; and Duplaa, C. et al. (1993) Anal. Biochem. 229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in an ELISA format where the oligomer of interest is presented in various dilutions and a
30 spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as targets in a microarray. The

microarray can be used to monitor the expression level of large numbers of genes simultaneously and to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, and to develop and monitor the activities of therapeutic agents.

In one embodiment, the microarray is prepared and used according to methods known in the art. (See, e.g., Chee et al. (1995) PCT application WO95/11995; Lockhart, D. J. et al. (1996) Nat. Biotech. 14:1675-1680; and Schena, M. et al. (1996) Proc. Natl. Acad. Sci. 93:10614-10619.)

The microarray is preferably composed of a large number of unique single-stranded nucleic acid sequences, usually either synthetic antisense oligonucleotides or fragments of cDNAs. The oligonucleotides are preferably about 6 to 60 nucleotides in length, more preferably about 15 to 30 nucleotides in length, and most preferably about 20 to 25 nucleotides in length. It may be preferable to use oligonucleotides which are about 7 to 10 nucleotides in length. The microarray may contain oligonucleotides which cover the known 5' or 3' sequence, sequential oligonucleotides which cover the full length sequence, or unique oligonucleotides selected from particular areas along the length of the sequence. Polynucleotides used in the microarray may be oligonucleotides specific to a gene or genes of interest. Oligonucleotides can also be specific to one or more unidentified cDNAs associated with a particular cell type or tissue type. It may be appropriate to use pairs of oligonucleotides on a microarray. The first oligonucleotide in each pair differs from the second oligonucleotide by one nucleotide. This nucleotide is preferably located in the center of the sequence. The second oligonucleotide serves as a control. The number of oligonucleotide pairs may range from about 2 to 1,000,000.

In order to produce oligonucleotides for use on a microarray, the gene of interest is examined using a computer algorithm which starts at the 5' end, or, more preferably, at the 3' end of the nucleotide sequence. The algorithm identifies oligomers of defined length that are unique to the gene, have a GC content within a range suitable for hybridization, and lack secondary structure that may interfere with hybridization. In one aspect, the oligomers may be synthesized on a substrate using a light-directed chemical process. (See, e.g., Chee et al., supra.) The substrate may be any suitable solid support, e.g., paper, nylon, any other type of membrane, or a filter, chip, or glass slide.

In another aspect, the oligonucleotides may be synthesized on the surface of the substrate using a chemical coupling procedure and an ink jet application apparatus. (See, e.g., Baldeschweiler et al. (1995) PCT application WO95/251116.) An array analogous to a dot or slot blot (HYBRIDOT® apparatus, GIBCO/BRL) may be used to arrange and link
5 cDNA fragments or oligonucleotides to the surface of a substrate using a vacuum system or thermal, UV, mechanical, or chemical bonding procedures. An array may also be produced by hand or by using available devices, materials, and machines, e.g. Brinkmann® multichannel pipettors or robotic instruments. The array may contain from 2 to 1,000,000 or any other feasible number of oligonucleotides.

10 In order to conduct sample analysis using the microarrays, polynucleotides are extracted from a sample. The sample may be obtained from any bodily fluid, e.g., blood, urine, saliva, phlegm, gastric juices, cultured cells, biopsies, or other tissue preparations. To produce probes, the polynucleotides extracted from the sample are used to produce nucleic acid sequences complementary to the nucleic acids on the microarray. If the
15 microarray contains cDNAs, antisense RNAs (aRNAs) are appropriate probes. Therefore, in one aspect, mRNA is reverse-transcribed to cDNA. The cDNA, in the presence of fluorescent label, is used to produce fragment or oligonucleotide aRNA probes. The fluorescently labeled probes are incubated with the microarray so that the probes hybridize to the microarray oligonucleotides. Nucleic acid sequences used as probes can include
20 polynucleotides, fragments, and complementary or antisense sequences produced using restriction enzymes, PCR, or other methods known in the art.

Hybridization conditions can be adjusted so that hybridization occurs with varying degrees of complementarity. A scanner can be used to determine the levels and patterns of fluorescence after removal of any nonhybridized probes. The degree of complementarity
25 and the relative abundance of each oligonucleotide sequence on the microarray can be assessed through analysis of the scanned images. A detection system may be used to measure the absence, presence, or level of hybridization for any of the sequences. (See, e.g., Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. 94:2150-2155.)

In another embodiment of the invention, nucleic acid sequences encoding HUPM
30 may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human

artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Price, C.M. (1993) *Blood Rev.* 7:127-134; and Trask, B.J. (1991) *Trends Genet.* 7:149-154.)

5 Fluorescent in situ hybridization (FISH) may be correlated with other physical chromosome mapping techniques and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, R.A. (ed.) Molecular Biology and Biotechnology, VCH Publishers New York, NY, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) site. Correlation between
10 the location of the gene encoding HUPM on a physical chromosomal map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder. The nucleotide sequences of the invention may be used to detect differences in gene sequences among normal, carrier, and affected individuals.

In situ hybridization of chromosomal preparations and physical mapping
15 techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the number or arm of a particular human chromosome is not known. New sequences can be assigned to chromosomal arms by physical mapping. This provides valuable information to
20 investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the disease or syndrome has been crudely localized by genetic linkage to a particular genomic region, e.g., AT to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) *Nature* 336:577-580.) The nucleotide sequence of the subject invention
25 may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

 In another embodiment of the invention, HUPM, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening
30 may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between HUPM and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate, such as plastic pins or some
5 other surface. The test compounds are reacted with HUPM, or fragments thereof, and washed. Bound HUPM is then detected by methods well known in the art. Purified HUPM can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

10 In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding HUPM specifically compete with a test compound for binding HUPM. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with HUPM.

In additional embodiments, the nucleotide sequences which encode HUPM may be
15 used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

The examples below are provided to illustrate the subject invention and are not
20 included for the purpose of limiting the invention.

EXAMPLES

For purposes of example, the preparation and sequencing of the PROSTUT03
25 cDNA library, from which Incyte Clone 789927 was isolated, is described. Preparation and sequencing of cDNAs in libraries in the LIFESEQ™ database have varied over time, and the gradual changes involved use of kits, plasmids, and machinery available at the particular time the library was made and analyzed.

I. PROSTUT03 cDNA Library Construction

30 The PROSTUT03 cDNA library was constructed from prostate tumor tissue removed from a 76-year-old Caucasian male by radical prostatectomy. The pathology report indicated grade 3 (of 4) adenocarcinoma (Gleason grade 3+3) in the periphery of the

prostate. Perineural invasion was present as was involvement of periprostatic tissue. Non-tumorous portions of the prostate exhibited adenofibromatous hyperplasia. The patient had elevated levels of prostate specific antigen (PSA). Pelvic lymph nodes were negative for tumor. A prior stomach ulcer and atherosclerosis were reported in the patient history.

5 The frozen tissue was homogenized and lysed using a Brinkmann Homogenizer Polytron-PT 3000 (Brinkmann Instruments, Inc. Westbury NY) in guanidinium isothiocyanate solution. The lysate was extracted once with acid phenol at pH 4.0 per Stratagene's RNA isolation protocol (Stratagene Inc.) and once with phenol chloroform at pH 4.0. The RNA was precipitated using 0.3 M sodium acetate and 2.5 volumes of
10 ethanol, resuspended in RNase-free water, and treated with DNase at 37°C. RNA extraction and precipitation were repeated as before. The mRNA was isolated with the Qiagen Oligotex kit (QIAGEN, Inc., Chatsworth, CA) and used to construct the cDNA library.

 The RNA was handled according to the recommended protocols in the SuperScript
15 Plasmid System (catalog #18248-013; Gibco/BRL). PROSTUT03 cDNAs were fractionated on a Sepharose CL4B column (catalog #275105, Pharmacia), and those cDNAs exceeding 400 bp were ligated into pSport I. The plasmid pSport I was subsequently transformed into DH5a™ competent cells (Cat. #18258-012, Gibco/BRL).

20 **II. Isolation and Sequencing of cDNA Clones**

 Plasmid DNA was released from the cells and purified using the REAL Prep 96 plasmid kit (Catalog #26173; QIAGEN, Inc.). The recommended protocol was employed except for the following changes: 1) the bacteria were cultured in 1 ml of sterile Terrific Broth (Catalog #22711, Gibco/BRL) with carbenicillin at 25 mg/L and glycerol at 0.4%;
25 2) after inoculation, the cultures were incubated for 19 hours and at the end of incubation, the cells were lysed with 0.3 ml of lysis buffer; and 3) following isopropanol precipitation, the plasmid DNA pellet was resuspended in 0.1 ml of distilled water. After the last step in the protocol, samples were transferred to a 96-well block for storage at 4° C.

 The cDNAs were sequenced by the method of Sanger, et al. (1975, J. Mol. Biol.
30 94:441f), using a Hamilton Micro Lab 2200 (Hamilton, Reno, NV) in combination with Peltier Thermal Cyclers (PTC200 from MJ Research, Watertown, MA) and Applied Biosystems 377 DNA Sequencing Systems.

III. Homology Searching of cDNA Clones and Their Deduced Proteins

The nucleotide sequences and/or amino acid sequences of the Sequence Listing were used to query sequences in the GenBank, SwissProt, BLOCKS, and Pima II databases. These databases, which contain previously identified and annotated sequences, were searched for regions of homology using BLAST, which stands for Basic Local Alignment Search Tool. (Altschul, S.F. (1993) J. Mol. Evol 36:290-300; and Altschul, et al. (1990) J. Mol. Biol. 215:403-410.)

BLAST produced alignments of both nucleotide and amino acid sequences to determine sequence similarity. Because of the local nature of the alignments, BLAST was especially useful in determining exact matches or in identifying homologs which may be of prokaryotic (bacterial) or eukaryotic (animal, fungal, or plant) origin. Other algorithms could have been used when dealing with primary sequence patterns and secondary structure gap penalties. (See, e.g., Smith, T. et al. (1992) Protein Engineering 5:35-51.) The sequences disclosed in this application have lengths of at least 49 nucleotides, and no more than 12% uncalled bases (where N can be A, C, G, or T).

The BLAST approach searched for matches between a query sequence and a database sequence. BLAST evaluated the statistical significance of any matches found, and reported only those matches that satisfy the user-selected threshold of significance. In this application, threshold was set at 10^{-25} for nucleotides and 10^{-10} for peptides.

Incyte nucleotide sequences were searched against the GenBank databases for primate (pri), rodent (rod), and other mammalian sequences (mam); and deduced amino acid sequences from the same clones were then searched against GenBank functional protein databases, mammalian (mamp), vertebrate (vrtp), and eukaryote (eukp) for homology.

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- 5 The sequences disclosed in this application have lengths of at least 49 nucleotides and have no more than 12% uncalled bases (where N is recorded rather than A, C, G, or T).

The BLAST approach searched for matches between a query sequence and a database sequence. BLAST evaluated the statistical significance of any matches found, and reported only those matches that satisfy the user-selected threshold of significance. In
10 this application, threshold was set at 10^{-25} for nucleotides and 10^{-8} for peptides.

Incyte nucleotide sequences were searched against the GenBank databases for primate (pri), rodent (rod), and other mammalian sequences (mam), and deduced amino acid sequences from the same clones were then searched against GenBank functional protein databases, mammalian (mamp), vertebrate (vrtp), and eukaryote (eukp), for
15 homology.

IV. Northern Analysis

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See,
20 e.g., Sambrook, *supra*, ch. 7; and Ausubel, F.M. et al. *supra*, ch. 4 and 16.)

Analogous computer techniques applying BLAST are used to search for identical or related molecules in nucleotide databases such as GenBank or LIFESEQ™ database (Incyte Pharmaceuticals). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to
25 determine whether any particular match is categorized as exact or homologous.

The basis of the search is the product score, which is defined as:

$$\frac{\% \text{ sequence identity} \times \% \text{ maximum BLAST score}}{100}$$

- 30 The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. For example, with a product score of 40, the match will be exact within a 1% to 2% error, and, with a product score of 70, the match will be

exact. Homologous molecules are usually identified by selecting those which show product scores between 15 and 40, although lower scores may identify related molecules.

The results of northern analysis are reported as a list of libraries in which the transcript encoding HUPM occurs. Abundance and percent abundance are also reported.

- 5 Abundance directly reflects the number of times a particular transcript is represented in a cDNA library, and percent abundance is abundance divided by the total number of sequences examined in the cDNA library.

V. Extension of HUPM Encoding Polynucleotides

- The sequence of one of the polynucleotides of the present invention was used to
 10 design oligonucleotide primers for extending a partial nucleotide sequence to full length. One primer was synthesized to initiate extension of an antisense polynucleotide, and the other was synthesized to initiate extension of a sense polynucleotide. Primers were used to facilitate the extension of the known sequence "outward" generating amplicons containing new unknown nucleotide sequence for the region of interest. The initial primers were
 15 designed from the cDNA using OLIGO 4.06 (National Biosciences, Plymouth, MN), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

- 20 Selected human cDNA libraries (GIBCO/BRL) were used to extend the sequence. If more than one extension is necessary or desired, additional sets of primers are designed to further extend the known region.

- High fidelity amplification was obtained by following the instructions for the XL-PCR kit (Perkin Elmer) and thoroughly mixing the enzyme and reaction mix. PCR was
 25 performed using the Peltier Thermal Cycler (PTC200; M.J. Research, Watertown, MA), beginning with 40 pmol of each primer and the recommended concentrations of all other components of the kit, with the following parameters:

- | | | |
|----|--------|--|
| | Step 1 | 94° C for 1 min (initial denaturation) |
| | Step 2 | 65° C for 1 min |
| 30 | Step 3 | 68° C for 6 min |
| | Step 4 | 94° C for 15 sec |
| | Step 5 | 65° C for 1 min |
| | Step 6 | 68° C for 7 min |
| | Step 7 | Repeat steps 4 through 6 for an additional 15 cycles |

- | | | |
|---|---------|---|
| | Step 8 | 94° C for 15 sec |
| | Step 9 | 65° C for 1 min |
| | Step 10 | 68° C for 7:15 min |
| | Step 11 | Repeat steps 8 through 10 for an additional 12 cycles |
| 5 | Step 12 | 72° C for 8 min |
| | Step 13 | 4° C (and holding) |

A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a low concentration (about 0.6% to 0.8%) agarose mini-gel to determine which reactions
 10 were successful in extending the sequence. Bands thought to contain the largest products were excised from the gel, purified using QIAQuick™ (QIAGEN Inc., Chatsworth, CA), and trimmed of overhangs using Klenow enzyme to facilitate religation and cloning.

After ethanol precipitation, the products were redissolved in 13 μ l of ligation buffer, 1 μ l T4-DNA ligase (15 units) and 1 μ l T4 polynucleotide kinase were added, and
 15 the mixture was incubated at room temperature for 2 to 3 hours, or overnight at 16° C. Competent *E. coli* cells (in 40 μ l of appropriate media) were transformed with 3 μ l of ligation mixture and cultured in 80 μ l of SOC medium. (See, e.g., Sambrook, *supra*, Appendix A, p. 2.) After incubation for one hour at 37° C, the *E. coli* mixture was plated on Luria Bertani (LB) agar (See, e.g., Sambrook, *supra*, Appendix A, p. 1) containing 2x
 20 Carb. The following day, several colonies were randomly picked from each plate and cultured in 150 μ l of liquid LB/2x Carb medium placed in an individual well of an appropriate commercially-available sterile 96-well microtiter plate. The following day, 5 μ l of each overnight culture was transferred into a non-sterile 96-well plate and, after dilution 1:10 with water, 5 μ l from each sample was transferred into a PCR array.

For PCR amplification, 18 μ l of concentrated PCR reaction mix (3.3x) containing
 25 4 units of rTth DNA polymerase, a vector primer, and one or both of the gene specific primers used for the extension reaction were added to each well. Amplification was performed using the following conditions:

- | | | |
|----|--------|--|
| | Step 1 | 94° C for 60 sec |
| 30 | Step 2 | 94° C for 20 sec |
| | Step 3 | 55° C for 30 sec |
| | Step 4 | 72° C for 90 sec |
| | Step 5 | Repeat steps 2 through 4 for an additional 29 cycles |
| | Step 6 | 72° C for 180 sec |
| 35 | Step 7 | 4° C (and holding) |

Aliquots of the PCR reactions were run on agarose gels together with molecular

weight markers. The sizes of the PCR products were compared to the original partial cDNAs, and appropriate clones were selected, ligated into plasmid, and sequenced.

The nucleotide sequences of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24 are used to obtain 5' regulatory sequences using the procedure above, oligonucleotides designed for 5' extension, and an appropriate genomic library.

VI. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [γ - 32 P] adenosine triphosphate (Amersham, Chicago, IL), and T4 polynucleotide kinase (DuPont NEN[®], Boston, MA). The labeled oligonucleotides are substantially purified using a Sephadex G-25 superfine resin column (Pharmacia & Upjohn, Kalamazoo, MI). An aliquot containing 10^7 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN, Boston, MA).

The DNA from each digest is fractionated on a 0.7 percent agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham, NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under increasingly stringent conditions up to 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. After XOMAT AR[™] film (Kodak, Rochester, NY) is exposed to the blots to film for several hours, hybridization patterns are compared visually.

VII. Microarrays

To produce oligonucleotides for a microarray, one of the nucleotide sequences of the present invention is examined using a computer algorithm which starts at the 3' end of

the nucleotide sequence. For each, the algorithm identifies oligomers of defined length that are unique to the nucleic acid sequence, have a GC content within a range suitable for hybridization, and lack secondary structure that would interfere with hybridization. The algorithm identifies approximately 20 oligonucleotides corresponding to each nucleic acid
5 sequence. For each sequence-specific oligonucleotide, a pair of oligonucleotides is synthesized in which the first oligonucleotide differs from the second oligonucleotide by one nucleotide in the center of the sequence. The oligonucleotide pairs can be arranged on a substrate, e.g. a silicon chip, using a light-directed chemical process. (See, e.g., Chee, supra.)

10 In the alternative, a chemical coupling procedure and an ink jet device can be used to synthesize oligomers on the surface of a substrate. (See, e.g., Baldeschweiler, supra.) An array analogous to a dot or slot blot may also be used to arrange and link fragments or oligonucleotides to the surface of a substrate using or thermal, UV, mechanical, or chemical bonding procedures, or a vacuum system. A typical array may be produced by
15 hand or using available methods and machines and contain any appropriate number of elements. After hybridization, nonhybridized probes are removed and a scanner used to determine the levels and patterns of fluorescence. The degree of complementarity and the relative abundance of each oligonucleotide sequence on the microarray may be assessed through analysis of the scanned images.

20 VIII. Complementary Polynucleotides

Sequences complementary to the HUPM-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring HUPM. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments.
25 Appropriate oligonucleotides are designed using Oligo 4.06 software and the coding sequence of HUPM. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the HUPM-encoding transcript.

30 IX. Expression of HUPM

Expression of HUPM is accomplished by subcloning the cDNA into an appropriate vector and transforming the vector into host cells. This vector contains an

appropriate promoter, e.g., β -galactosidase upstream of the cloning site, operably associated with the cDNA of interest. (See, e.g., Sambrook, supra, pp. 404-433; and Rosenberg, M. et al. (1983) *Methods Enzymol.* 101:123-138.)

Induction of an isolated, transformed bacterial strain with isopropyl beta-D-
5 thiogalactopyranoside (IPTG) using standard methods produces a fusion protein which consists of the first 8 residues of β -galactosidase, about 5 to 15 residues of linker, and the full length protein. The signal residues direct the secretion of HUPM into bacterial growth media which can be used directly in the following assay for activity.

X. Demonstration of HUPM Activity

10 Serine protease activity of HUPM is measured by the hydrolysis of various peptide thiobenzyl ester substrates. The substrates are chosen to represent the different SP types (chymase, trypase, aspase, etc.). Assays are performed at room temperature ($\sim 25^{\circ}\text{C}$) and contain an aliquot of HUPM and the appropriate substrate in HEPES buffer, pH 7.5 containing 0.01M CaCl_2 and 8% dimethylsulfoxide. The reaction also contains 0.34 mM
15 dithiopyridine which reacts with the thiobenzyl group that is released during hydrolysis and converts it to thiopyridone. The reaction is carried out in an optical cuvette,, and the generation of thiopyridone is measured in a spectrophotometer by the absorption produced at 324 nm. The amount of thiopyridone produced in the reaction is proportional to the activity of HUPM.

20 XI. Production of HUPM Specific Antibodies

HUPM substantially purified using PAGE electrophoresis (see, e.g., Harrington, M.G. (1990) *Methods Enzymol.* 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols. The HUPM amino acid sequence is analyzed using DNASTAR software (DNASTAR Inc) to determine
25 regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel et al. supra, ch. 11.)

Typically, the oligopeptides are 15 residues in length, and are synthesized using
30 an Applied Biosystems Peptide Synthesizer Model 431A using fmoc-chemistry and coupled to KLH (Sigma, St. Louis, MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester to increase immunogenicity. (See, e.g., Ausubel et al. supra.)

Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide activity, for example, by binding the peptide to plastic, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

5 **XII. Purification of Naturally Occurring HUPM Using Specific Antibodies**

Naturally occurring or recombinant HUPM is substantially purified by immunoaffinity chromatography using antibodies specific for HUPM. An immunoaffinity column is constructed by covalently coupling anti-HUPM antibody to an activated chromatographic resin, such as CNBr-activated Sepharose (Pharmacia & Upjohn). After
10 the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing HUPM are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of HUPM (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/HUPM binding (e.g., a buffer of pH 2 to pH 3, or a high
15 concentration of a chaotrope, such as urea or thiocyanate ion), and HUPM is collected.

XIII. Identification of Molecules Which Interact with HUPM

HUPM, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton et al. (1973) Biochem. J. 133:529.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the
20 labeled HUPM, washed, and any wells with labeled HUPM complex are assayed. Data obtained using different concentrations of HUPM are used to calculate values for the number, affinity, and association of HUPM with the candidate molecules.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and
25 spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following
30 claims.

What is claimed is:

1. A substantially purified human protease molecule (HUPM) comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:2,
5 SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, and SEQ ID NO:12, and fragments thereof.
2. A substantially purified variant of HUPM having at least 90% amino acid
10 identity to the amino acid sequence of claim 1.
3. An isolated and purified polynucleotide sequence encoding the HUPM of claim 1.
- 15 4. An isolated and purified polynucleotide variant having at least 90% polynucleotide identity to the polynucleotide sequence of claim 3.
5. A composition comprising the polynucleotide sequence of claim 3.
- 20 6. An isolated and purified polynucleotide sequence which hybridizes under stringent conditions to the polynucleotide sequence of claim 3.
7. An isolated and purified polynucleotide sequence which is complementary to the polynucleotide sequence of claim 3.
25
8. An isolated and purified polynucleotide sequence comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, SEQ ID NO:16, SEQ ID NO:17, SEQ ID NO:18, SEQ ID NO:19, SEQ ID NO:20, SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23, and SEQ ID NO:24,
30 and fragments thereof.
9. An isolated and purified polynucleotide variant having at least 90%

polynucleotide identity to the polynucleotide sequence of claim 8.

10. An isolated and purified polynucleotide sequence which is complementary to the polynucleotide sequence of claim 8.

5

11. An expression vector containing at least a fragment of the polynucleotide sequence of claim 3.

12. A host cell containing the expression vector of claim 11.

10

13. A method for producing a polypeptide comprising the amino acid sequence of SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:4, SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, or fragments thereof, the method comprising the steps of:

15

a) culturing the host cell of claim 12 under conditions suitable for the expression of the polypeptide; and

b) recovering the polypeptide from the host cell culture.

14. A pharmaceutical composition comprising the HUPM of claim 1 in conjunction with a suitable pharmaceutical carrier.

20

15. A purified antibody which specifically binds to the HUPM of claim 1.

16. A purified agonist of the HUPM of claim 1.

25

17. A purified antagonist of the HUPM of claim 1.

18. A method for treating or preventing a cell proliferative disorder, the method comprising administering to a subject in need of such treatment an effective amount of the pharmaceutical composition of claim 14.

30

19. A method for treating or preventing an immune disorder, the method

comprising administering to a subject in need of such treatment an effective amount of the pharmaceutical composition of claim 14.

20. A method for treating or preventing a cell proliferative disorder, the method
5 comprising administering to a subject in need of such treatment an effective amount of the purified antagonist of claim 17.

21. A method for treating or preventing an immune disorder, the method
comprising administering to a subject in need of such treatment an effective amount of the
10 purified antagonist of claim 17.

22. A method for detecting a polynucleotide encoding HUPM in a biological sample containing nucleic acids, the method comprising the steps of:

(a) hybridizing the polynucleotide of claim 7 to at least one of the
15 nucleic acids in the biological sample, thereby forming a hybridization complex;
and

(b) detecting the hybridization complex, wherein the presence of the
hybridization complex correlates with the presence of a polynucleotide encoding
HUPM in the biological sample.

20

23. The method of claim 22 wherein the nucleic acids of the biological sample are amplified by the polymerase chain reaction prior to hybridization.

SEQUENCE LISTING

<110> INCYTE PHARMACEUTICALS, INC.

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HILLMAN, Jennifer L.

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GUEGLER, Karl J.

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TANG, Y. Tom

SHAH, Purvi

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<223> 2170967

<400> 9

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	20	25	30
Asp Met Thr Lys	Gly Leu Val Leu	Gly Ile Tyr Ser	Lys Glu Lys
	35	40	45
Glu Asp Asp Val	Pro Gln Phe Thr	Ser Ala Gly Glu	Asn Phe Asp
	50	55	60
Lys Leu Leu Ala	Gly Lys Leu Arg	Glu Thr Leu Asn	Ile Ser Gly
	65	70	75
Pro Pro Leu Lys	Ala Gly Lys Thr	Arg Thr Phe Tyr	Gly Leu His
	80	85	90
Gln Asp Phe Pro	Ser Val Val Leu	Val Gly Leu Gly	Lys Lys Ala
	95	100	105
Ala Gly Ile Asp	Glu Gln Glu Asn	Trp His Glu Gly	Lys Glu Asn
	110	115	120
Ile Arg Ala Ala	Val Ala Ala Gly	Cys Arg Gln Ile	Gln Asp Leu
	125	130	135
Glu Leu Ser Ser	Val Glu Val Asp	Pro Cys Gly Asp	Ala Gln Ala
	140	145	150

Ala	Ala	Glu	Gly	Ala	Val	Leu	Gly	Leu	Tyr	Glu	Tyr	Asp	Asp	Leu
				155						160				165
Lys	Gln	Lys	Lys	Lys	Met	Ala	Val	Ser	Ala	Lys	Leu	Tyr	Gly	Ser
				170						175				180
Gly	Asp	Gln	Glu	Ala	Trp	Gln	Lys	Gly	Val	Leu	Phe	Ala	Ser	Gly
				185						190				195
Gln	Asn	Leu	Ala	Arg	Gln	Leu	Met	Glu	Thr	Pro	Ala	Asn	Glu	Met
				200						205				210
Thr	Pro	Thr	Arg	Phe	Ala	Glu	Ile	Ile	Glu	Lys	Asn	Leu	Lys	Ser
				215						220				225
Ala	Ser	Ser	Lys	Thr	Glu	Val	His	Ile	Arg	Pro	Lys	Ser	Trp	Ile
				230						235				240
Glu	Glu	Gln	Ala	Met	Gly	Ser	Phe	Leu	Ser	Val	Ala	Lys	Gly	Ser
				245						250				255
Asp	Glu	Pro	Pro	Val	Phe	Leu	Glu	Ile	His	Tyr	Lys	Gly	Ser	Pro
				260						265				270
Asn	Ala	Asn	Glu	Pro	Pro	Leu	Val	Phe	Val	Gly	Lys	Gly	Ile	Thr
				275						280				285
Phe	Asp	Ser	Gly	Gly	Ile	Ser	Ile	Lys	Ala	Ser	Ala	Asn	Met	Asp
				290						295				300
Leu	Met	Arg	Ala	Asp	Met	Gly	Gly	Ala	Ala	Thr	Ile	Cys	Ser	Ala
				305						310				315
Ile	Val	Ser	Ala	Ala	Lys	Leu	Asn	Leu	Pro	Ile	Asn	Ile	Ile	Gly
				320						325				330
Leu	Ala	Pro	Leu	Cys	Glu	Asn	Met	Pro	Ser	Gly	Lys	Ala	Asn	Lys
				335						340				345
Pro	Gly	Asp	Val	Val	Arg	Ala	Lys	Asn	Gly	Lys	Thr	Ile	Gln	Val
				350						355				360
Asp	Asn	Thr	Asp	Ala	Glu	Gly	Arg	Leu	Ile	Leu	Ala	Asp	Ala	Leu
				365						370				375
Cys	Tyr	Ala	His	Thr	Phe	Asn	Pro	Lys	Val	Ile	Leu	Asn	Ala	Ala
				380						385				390
Thr	Leu	Thr	Gly	Ala	Met	Asp	Val	Ala	Leu	Gly	Ser	Gly	Ala	Thr
				395						400				405
Gly	Val	Phe	Thr	Asn	Ser	Ser	Trp	Leu	Trp	Asn	Lys	Leu	Phe	Glu
				410						415				420
Ala	Ser	Ile	Glu	Thr	Gly	Asp	Arg	Val	Trp	Arg	Met	Pro	Leu	Phe
				425						430				435
Glu	His	Tyr	Thr	Arg	Gln	Val	Val	Asp	Cys	Gln	Leu	Ala	Asp	Val
				440						445				450
Asn	Asn	Ile	Gly	Lys	Tyr	Arg	Ser	Ala	Gly	Ala	Cys	Thr	Ala	Ala
				455						460				465
Ala	Phe	Leu	Lys	Glu	Phe	Val	Thr	His	Pro	Lys	Trp	Ala	His	Leu
				470						475				480
Asp	Ile	Ala	Gly	Val	Met	Thr	Asn	Lys	Asp	Glu	Val	Pro	Tyr	Leu
				485						490				495
Arg	Lys	Gly	Met	Thr	Gly	Arg	Pro	Thr	Arg	Thr	Leu	Ile	Glu	Phe
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Leu	Leu	Arg	Phe	Ser	Gln	Asp	Asn	Ala						
				515										

<210> 10

<211> 327

<212> PRT

<213> Homo sapiens

<220> -

<223> 2484218

<400> 10

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Gly Ser Ser Gly Met Glu Val Asp Ala Ala Val Val Pro Ser Val
          20           25           30
Met Ala Cys Gly Val Thr Gly Ser Val Ser Val Ala Leu His Pro
          35           40           45
Leu Val Ile Leu Asn Ile Ser Asp His Trp Ile Arg Met Arg Ser
          50           55           60
Gln Glu Gly Arg Pro Val Gln Val Ile Gly Ala Leu Ile Gly Lys
          65           70           75
Gln Glu Gly Arg Asn Ile Glu Val Met Asn Ser Phe Glu Leu Leu
          80           85           90
Ser His Thr Val Glu Glu Lys Ile Ile Ile Asp Lys Glu Tyr Tyr
          95          100          105
Tyr Thr Lys Glu Glu Gln Phe Lys Gln Val Phe Lys Glu Leu Glu
          110          115          120
Phe Leu Gly Trp Tyr Thr Thr Gly Gly Pro Pro Asp Pro Ser Asp
          125          130          135
Ile His Val His Lys Gln Val Cys Glu Ile Ile Glu Ser Pro Leu
          140          145          150
Phe Leu Lys Leu Asn Pro Met Thr Lys His Thr Asp Leu Pro Val
          155          160          165
Ser Val Phe Glu Ser Val Ile Asp Ile Ile Asn Gly Glu Ala Thr
          170          175          180
Met Leu Phe Ala Glu Leu Thr Tyr Thr Leu Ala Thr Glu Glu Ala
          185          190          195
Glu Arg Ile Gly Val Asp His Val Ala Arg Met Thr Ala Thr Gly
          200          205          210
Ser Gly Glu Asn Ser Thr Val Ala Glu His Leu Ile Ala Gln His
          215          220          225
Ser Ala Ile Lys Met Leu His Ser Arg Val Lys Leu Ile Leu Glu
          230          235          240
Tyr Val Lys Ala Ser Glu Ala Gly Glu Val Pro Phe Asn His Glu
          245          250          255
Ile Leu Arg Glu Ala Tyr Ala Leu Cys His Cys Leu Pro Val Leu
          260          265          270
Ser Thr Asp Lys Phe Lys Thr Asp Phe Tyr Asp Gln Cys Asn Asp
          275          280          285
Val Gly Leu Met Ala Tyr Leu Gly Thr Ile Thr Lys Thr Cys Asn
          290          295          300
Thr Met Asn Gln Phe Val Asn Lys Phe Asn Val Leu Tyr Asp Arg
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Gln Gly Ile Gly Arg Arg Met Arg Gly Leu Phe Phe
          320          325

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<210> 11

<211> 458
 <212> PRT
 <213> Homo sapiens

<220> -
 <223> 2680548

<400> 11

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Leu	Thr	Pro	Asp	Leu	Arg	Ala	Leu	Leu	Thr	Ser	Gly	Thr	Ser	Asp	35	40	45	
Pro	Arg	Ala	Arg	Val	Thr	Tyr	Gly	Thr	Pro	Ser	Leu	Trp	Ala	Arg	50	55	60	
Leu	Ser	Val	Gly	Val	Thr	Glu	Pro	Arg	Ala	Cys	Leu	Thr	Ser	Gly	65	70	75	
Thr	Pro	Gly	Pro	Arg	Ala	Gln	Leu	Thr	Ala	Val	Thr	Pro	Asp	Thr	80	85	90	
Arg	Thr	Arg	Glu	Ala	Ser	Glu	Asn	Ser	Gly	Thr	Arg	Ser	Arg	Ala	95	100	105	
Trp	Leu	Ala	Val	Ala	Leu	Gly	Ala	Gly	Gly	Ala	Val	Leu	Leu	Leu	110	115	120	
Leu	Trp	Gly	Gly	Gly	Arg	Gly	Pro	Pro	Ala	Val	Leu	Ala	Ala	Val	125	130	135	
Pro	Ser	Pro	Pro	Pro	Ala	Ser	Pro	Arg	Ser	Gln	Tyr	Asn	Phe	Ile	140	145	150	
Ala	Asp	Val	Val	Glu	Lys	Thr	Ala	Pro	Ala	Val	Val	Tyr	Ile	Glu	155	160	165	
Ile	Leu	Asp	Arg	His	Pro	Phe	Leu	Gly	Arg	Glu	Val	Pro	Ile	Ser	170	175	180	
Asn	Gly	Ser	Gly	Phe	Val	Val	Ala	Ala	Asp	Gly	Leu	Ile	Val	Thr	185	190	195	
Asn	Ala	His	Val	Val	Ala	Asp	Arg	Arg	Arg	Val	Arg	Val	Arg	Leu	200	205	210	
Leu	Ser	Gly	Asp	Thr	Tyr	Glu	Ala	Val	Val	Thr	Ala	Val	Asp	Pro	215	220	225	
Val	Ala	Asp	Ile	Ala	Thr	Leu	Arg	Ile	Gln	Thr	Lys	Glu	Pro	Leu	230	235	240	
Pro	Thr	Leu	Pro	Leu	Gly	Arg	Ser	Ala	Asp	Val	Arg	Gln	Gly	Glu	245	250	255	
Phe	Val	Val	Ala	Met	Gly	Ser	Pro	Phe	Ala	Leu	Gln	Asn	Thr	Ile	260	265	270	
Thr	Ser	Gly	Ile	Val	Ser	Ser	Ala	Gln	Arg	Pro	Ala	Arg	Asp	Leu	275	280	285	
Gly	Leu	Pro	Gln	Thr	Asn	Val	Glu	Tyr	Ile	Gln	Thr	Asp	Ala	Ala	290	295	300	
Ile	Asp	Phe	Gly	Asn	Ser	Gly	Gly	Pro	Leu	Val	Asn	Leu	Asp	Gly	305	310	315	
Glu	Val	Ile	Gly	Val	Asn	Thr	Met	Lys	Val	Thr	Ala	Gly	Ile	Ser	320	325	330	
Phe	Ala	Ile	Pro	Ser	Asp	Arg	Leu	Arg	Glu	Phe	Leu	His	Arg	Gly	335	340	345	
Glu	Lys	Lys	Asn	Ser	Ser	Ser	Gly	Ile	Ser	Gly	Ser	Gln	Arg	Arg	350	355	360	

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Tyr Ile Gly Val Met Met Leu Thr Leu Ser Pro Ser Ile Leu Ala
      365                      370                      375
Glu Leu Gln Leu Arg Glu Pro Ser Phe Pro Asp Val Gln His Gly
      380                      385                      390
Val Leu Ile His Lys Val Ile Leu Gly Ser Pro Ala His Arg Ala
      395                      400                      405
Gly Leu Arg Pro Gly Asp Val Ile Leu Ala Ile Gly Glu Gln Met
      410                      415                      420
Val Gln Asn Ala Glu Asp Val Tyr Glu Ala Val Arg Thr Gln Ser
      425                      430                      435
Gln Leu Ala Val Gln Ile Arg Arg Gly Arg Glu Thr Leu Thr Leu
      440                      445                      450
Tyr Val Thr Pro Glu Val Thr Glu
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<210> 12
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<212> PRT
<213> Homo sapiens

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<220> -
<223> 2957969

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<400> 12
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Gln Phe Gln Ala Ser Ala Gly Asp Ser Asp Pro Gln Ser Arg Pro
      20           25           30
Leu Leu Leu Leu Gly Gln Leu His His Leu His Arg Val Pro Trp
      35           40           45
Ser His Val Arg Gly Lys Leu Gln Pro Arg Val Thr Glu Glu Leu
      50           55           60
Trp Gln Ala Ala Leu Ser Thr Leu Asn Pro Asn Pro Thr Asp Ser
      65           70           75
Cys Pro Leu Tyr Leu Asn Tyr Ala Thr Val Ala Ala Leu Pro Cys
      80           85           90
Arg Val Ser Arg His Asn Ser Pro Ser Ala Ala His Phe Ile Thr
      95          100          105
Arg Leu Val Arg Thr Cys Leu Pro Pro Gly Ala His Arg Cys Ile
     110          115          120
Val Met Val Cys Glu Gln Pro Glu Val Phe Ala Ser Ala Cys Ala
     125          130          135
Leu Ala Arg Ala Phe Pro Leu Phe Thr His Arg Ser Gly Ala Ser
     140          145          150
Arg Arg Leu Glu Lys Lys Thr Val Thr Val Glu Phe Phe Leu Val
     155          160          165
Gly Gln Asp Asn Gly Pro Val Glu Val Ser Thr Leu Gln Cys Leu
     170          175          180
Ala Asn Ala Thr Asp Gly Val Arg Leu Ala Ala Arg Ile Val Asp
     185          190          195
Thr Pro Cys Asn Glu Met Asn Thr Asp Thr Phe Leu Glu Glu Ile
     200          205          210
Asn Lys Val Gly Lys Glu Leu Gly Ile Ile Pro Thr Ile Ile Arg

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	215		220		225
Asp Glu Glu Leu Lys Thr Arg Gly Phe Gly Gly Ile Tyr Gly Val					
	230		235		240
Gly Lys Ala Ala Leu His Pro Pro Ala Leu Ala Val Leu Ser His					
	245		250		255
Thr Pro Asp Gly Ala Thr Gln Thr Ile Ala Trp Val Gly Lys Gly					
	260		265		270
Ile Val Tyr Asp Thr Gly Gly Leu Ser Ile Lys Gly Lys Thr Thr					
	275		280		285
Met Pro Gly Met Lys Arg Asp Cys Gly Gly Ala Ala Ala Val Leu					
	290		295		300
Gly Ala Phe Arg Ala Ala Ile Lys Gln Gly Phe Lys Asp Asn Leu					
	305		310		315
His Ala Val Phe Cys Leu Ala Glu Asn Ser Val Gly Pro Asn Ala					
	320		325		330
Thr Arg Pro Asp Asp Ile His Leu Leu Tyr Ser Gly Lys Thr Val					
	335		340		345
Glu Ile Asn Asn Thr Asp Ala Glu Gly Arg Leu Val Leu Ala Asp					
	350		355		360
Gly Val Ser Tyr Ala Cys Lys Asp Leu Gly Ala Asp Ile Ile Leu					
	365		370		375
Asp Met Ala Thr Leu Thr Gly Ala Gln Gly Ile Ala Thr Gly Lys					
	380		385		390
Tyr His Ala Ala Val Leu Thr Asn Ser Ala Glu Trp Glu Ala Ala					
	395		400		405
Cys Val Lys Ala Gly Arg Lys Cys Gly Asp Leu Val His Pro Leu					
	410		415		420
Val Tyr Cys Pro Glu Leu His Phe Ser Glu Phe Thr Ser Ala Val					
	425		430		435
Ala Asp Met Lys Asn Ser Val Ala Asp Arg Asp Asn Ser Pro Ser					
	440		445		450
Ser Cys Ala Gly Leu Phe Ile Ala Ser His Ile Gly Phe Asp Trp					
	455		460		465
Pro Gly Val Trp Val His Leu Asp Ile Ala Ala Pro Val His Ala					
	470		475		480
Gly Glu Arg Ala Thr Gly Phe Gly Val Ala Leu Leu Leu Ala Leu					
	485		490		495
Phe Gly Arg Ala Ser Glu Asp Pro Leu Leu Asn Leu Val Ser Pro					
	500		505		510
Leu Gly Cys Glu Val Asp Val Glu Glu Gly Asp Val Gly Arg Asp					
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Ser Lys Arg Arg Arg Leu Val					
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<210> 13
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 <212> DNA
 <213> Homo sapiens

<220> -
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<400> 13


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caggatggag tgcagtagtg tgatcttggc tcaactgcaac ccctgcctcc cgggttcaag 180
gaattctcct gccttagcct cctgagtagc tgagattaca ggccgccacc accacacctg 240
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cactgtgccc agccaagtta tatctctaaa gcaatgtgca aaaataaact gaacttgggt 420
tgattaggta tattcaacat ttgtcgggag agtagatgtt tcattttatt tcagtccctg 480
tgtaatttgt cttctctaatt gttaaatact atgtagaatg tgtctgtgta attttataga 540
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atgtttggag gttacccaaa tcttaccttt cctttttcta ttctagaatt tacatagtac 660
atgacgaagt taaggataaa gcttttgaac tagaactcag ctgggttggg gaatgtaagt 720
tatttttgta catttatttg ccttaggaat gatctgtacc acagctaatt tacaactgag 780
tgtcctttct aatataatga aagctaaagc aaatttacta ggttgtctaa tgaagggaag 840
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<212> DNA

<213> Homo sapiens

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aaataattgt tttaattatt ttactgtagc atagattcta ttacaaaaat gttgttttat 1080
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cctgaaccca	cctggccccc	tcagctctgt	cctgttaggc	caactgcctc	cttggcagcc	3000
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<212> DNA

<213> Homo sapiens

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<223> 789927

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gccgaactcg	ggcgttggtc	gtggcagggg	agcctgcgcc	tgtgggattc	ccacgtatgc	240
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agtgaacctta	gtgatccctc	cgggtggatg	gtccagtttg	gccagctgac	ttccatgcca	360
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ccccacaccc	tccaggaagt	tcaggctgcc	atcataaaca	actctatgtg	caaccacctc	660
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<212> DNA

<213> Homo sapiens

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<223> 877617

<400> 16

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<211> 2038
<212> DNA
<213> Homo sapiens

<220> -
<223> 1337018

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 <223> a or g or c or t, unknown, or other

<220> -

<223> 2082147

<400> 20

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<212> DNA

<213> Homo sapiens

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<212> DNA

<213> Homo sapiens

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<223> 2680548

<400> 23

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